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#### INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 7:

C12N 15/86, 15/35, 5/10, A61K 48/00

**A3** 

(11) International Publication Number:

WO 00/28061

(43) International Publication Date:

18 May 2000 (18.05.00)

(21) International Application Number:

PCT/US99/25694

(22) International Filing Date:

2 November 1999 (02.11.99)

(30) Priority Data:

60/107,114

5 November 1998 (05.11.98)

US

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(81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

#### Published

With international search report.

(88) Date of publication of the international search report:
3 August 2000 (03.08.00)

(54) Title: ADENO-ASSOCIATED VIRUS SEROTYPE I NUCLEIC ACID SEQUENCES, VECTORS AND HOST CELLS CONTAINING SAME

#### (57) Abstract

The nucleic acid sequences of adeno-associated virus (AAV) serotype 1 are provided, as are vectors and host cells containing these sequences and functional fragments thereof. Also provided are methods of delivering genes via AAV-1 derived vectors.

Express Mail No. ET033648243US

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## ADENO-ASSOCIATED VIRUS SEROTYPE I NUCLEIC ACID SEQUENCES, VECTORS AND HOST CELLS CONTAINING SAME

This work was supported by the National Institutes of Health, grant no. P30 DK47757-06 and PO1 HD32649-04. The US government may have certain rights in this invention.

### Field of the Invention

This invention relates generally to viral vector, and more particularly, to recombinant viral vectors useful for gene delivery.

### Background of the Invention

Adeno-associated viruses are small, single-stranded DNA viruses which require helper virus to facilitate efficient replication [K.I. Berns, Parvoviridae: the viruses and their replication, p. 1007-1041, in F.N. Fields et al., Fundamental virology, 3rd ed., vol. 2, (Lippencott-Raven Publishers, Philadelphia, PA) (1995)]. The 4.7 kb genome of AAV is characterized by two inverted terminal repeats (ITR) and two open reading frames which encode the Rep proteins and Cap proteins, respectively. The Rep reading frame encodes four proteins of molecular weight 78 kD, 68 kD, 52 kD and 40 kD. These proteins function mainly in regulating AAV replication and integration of the AAV into a host cell's chromosomes. The Cap reading frame encodes three structural proteins in molecular weight 85 kD (VP 1), 72 kD (VP2) and 61 kD (VP3) [Berns, cited above]. More than 80% of total proteins in AAV virion comprise VP3. The two ITRs are the only cis elements essential for AAV replication, packaging and integration. There are two conformations of AAV ITRs called "flip" and "flop". These differences in conformation originated from the replication model of adeno-associated virus which use the ITR to initiate and reinitiate the replication [R.O. Snyder et al., <u>J. Virol.</u>, <u>67</u>:6096-6104 (1993); K.I. Berns, Microbiological Reviews, 54:316-329 (1990)].

AAVs have been found in many animal species, including primates, canine, fowl and human [F.A. Murphy et al., "The Classification and Nomenclature of Viruses: Sixth Report of the International Committee on Taxonomy of Viruses",

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Archives of Virology, (Springer-Verlag, Vienna) (1995)]. In addition to five known primate AAVs (AAV-1 to AAV-5), AAV-6, another serotype closely related to AAV-2 and AAV-1 has also been isolated [E. A. Rutledge et al., <u>J. Virol., 72</u>:309-319 (1998)]. Among all known AAV serotypes, AAV-2 is perhaps the most well-characterized serotype, because its infectious clone was the first made [R.J. Samulski et al., <u>Proc. Natl. Acad. Sci. USA, 79</u>:2077-2081 (1982)]. Subsequently, the full sequences for AAV-3A, AAV-3B, AAV-4 and AAV-6 have also been determined [Rutledge, cited above; J.A.Chiorini et al., <u>J. Virol., 71</u>:6823-6833 (1997); S. Muramatsu et al., <u>Virol., 221</u>:208-217 (1996)]. Generally, all AAVs share more than 80% homology in nucleotide sequence.

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A number of unique properties make AAV a promising vector for human gene therapy [Muzyczka, Current Topics in Microbiology and Immunology, 158:97-129 (1992)]. Unlike other viral vectors, AAVs have not been shown to be associated with any known human disease and are generally not considered pathogenic. Wild type AAV is capable of integrating into host chromosomes in a site specific manner [R. M. 15 Kotin et al., Proc. Natl. Acad. Sci. USA, 87:2211-2215 (1990)- R.J. Samulski, EMBO J., 10(12):3941-3950 (1991)]. Recombinant AAV vectors can integrate into tissue cultured cells in chromosome 19 if the rep proteins are supplied in trans [C. Balague et al., J. Virol., 71:3299-3306 (1997); R. T. Surosky et al., J. Virol., 71:7951-7959 (1997)]. The integrated genomes of AAV have been shown to allow 20 long term gene expression in a number of tissues, including, muscle, liver, and brain [K. J. Fisher, Nature Med., 3(3):306-312 (1997); R. O. Snyder et al., Nature Genetics, 16:270-276 (1997); X. Xiao et al., Experimental Neurology, 144:113-124 (1997); Xiao, J. Virol., 70(11):8098-8108 (1996)].

AAV-2 has been shown to be present in about 80-90% of the human population. Earlier studies showed that neutralizing antibodies for AAV-2 are prevalent [W. P. Parks et al., <u>J. Virol.</u>, <u>2</u>:716-722 (1970)]. The presence of such antibodies may significantly decrease the usefulness of AAV vectors based on AAV-2 despite its other merits. What are needed in the art are vectors characterized by the

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advantages of AAV-2, including those described above, without the disadvantages, including the presence of neutralizing antibodies.

### Summary of the Invention

In one aspect, the invention provides an isolated AAV-1 nucleic acid molecule which is selected from among SEQ ID NO: 1, the strand complementary to SEQ ID NO: 1, and cDNA and RNA sequences complementary to SEQ ID NO: 1 and its complementary strand.

In another aspect, the present invention provides AAV ITR sequences, which include the 5' ITR sequences, nt 1 to 143 of SEQ ID NO: 1; the 3' ITR sequences, nt 4576 to 4718 of SEQ ID NO: 1, and fragments thereof.

In yet another aspect, the present invention provides a recombinant vector comprising an AAV-1 ITR and a selected transgene. Preferably, the vector comprises both the 5' and 3' AAV-1 ITRs between which the selected transgene is located.

In still another aspect, the invention provides a recombinant vector comprising an AAV-1 P5 promoter having the sequence of nt 236 to 299 of SEQ ID NO: 1 or a functional fragment thereof.

In a further aspect, the present invention provides a nucleic acid molecule encoding an AAV-1 rep coding region and an AAV-1 cap coding region.

In still another aspect, the present invention provides a host cell transduced with a

recombinant viral vector of the invention. The invention further provides a host cell stably transduced with an AAV-1 P5 promoter of the invention.

In still a further aspect, the present invention provides a pharmaceutical composition comprising a carrier and a vector of the invention.

In yet another aspect, the present invention provides a method for AAV-mediated delivery of a transgene to a host involving the step of delivering to a selected
host a recombinant viral vector comprising a selected transgene under the control of
sequences which direct expression thereof and an adeno-associated virus 1 (AAV-1)
virion.

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In another aspect, the invention provides a method for in vitro production of a selected gene product using a vector of the invention.

Other aspects and advantages of the invention will be readily apparent to one of skill in the art from the detailed description of the invention.

### 5 Brief Description of the Drawings

Figs. 1A-1C illustrate the alignment of nucleotides of AAV-1 [SEQ ID NO: 1], AAV-2 [SEQ ID NO: 18] and AAV-6 [SEQ ID NO: 19]. The alignment was done with MacVector 6.0. The full sequences of AAV-1 are shown in the top line. Nucleotides in AAV-2 and AAV-6 identical to AAV-1 are symbolized by "." and gaps by "-". Some of the conserved features among AAVs are marked in this figure. Note the 3' ITRs of AAV-1 and AAV-6 are shown in different orientations.

Fig. 2 illustrates the predicted secondary structure of AAV-1 ITR. The nucleotides in AAV-2 and AAV-6 are shown in italic and bold respectively.

Fig. 3A illustrates a hypothesis of how AAV-6 arose from the homologous recombination between AAV-1 and AAV-2. The major elements of AAV-1 are indicated in the graph. A region that is shared between AAV-1, AAV-2 and AAV-6 is shown in box with waved lines.

Fig. 3B is a detailed illustration of a 71 bp homologous region among AAV-1, AAV-2 and AAV-6. Nucleotides that differ among these serotypes are indicated by arrows.

Fig. 4A is a bar chart illustrating expression levels of human alpha 1 antitrypsin (a1AT) in serum following delivery of hAAT via recombinant AAV-1 and recombinant AAV-2 viruses.

Fig. 4B is a bar chart illustrating expression levels of erythropoietin (epo) in serum following delivery of the epo gene via recombinant AAV-1 and recombinant AAV-2 viruses.

Fig. 5A is a bar chart illustrating expression levels of  $\alpha 1AT$  in liver following delivery of  $\alpha 1AT$  as described in Example 7.

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Fig. 5B is a bar chart demonstrating expression levels of epo in liver following delivery of epo as described in Example 7.

Fig. 5C is a bar chart demonstrating neutralizing antibodies (NAB) directed to AAV-1 following delivery of a1AT or epo to liver as described in Example 7.

Fig. 5D is a bar chart demonstrating neutralizing antibodies (NAB) directed to AAV-2 following delivery of a1AT or epo to liver as described in Example 7.

Fig. 6A is a bar chart illustrating expression levels of  $\alpha 1$  AT in muscle following delivery of  $\alpha 1$  AT as described in Example 7.

Fig. 6B is a bar chart demonstrating expression levels of epo in muscle following delivery of epo as described in Example 7.

Fig. 6C is a bar chart demonstrating neutralizing antibodies (NAB) directed to AAV-1 following delivery of a1AT or epo to muscle as described in Example 7.

Fig. 6D is a bar chart demonstrating neutralizing antibodies (NAB) directed to AAV-2 following delivery of alAT or epo to muscle as described in Example 7.

### 15 Detailed Description of the Invention

The present invention provides novel nucleic acid sequences for an adeno-associated virus of serotype 1 (AAV-1). Also provided are fragments of these AAV-1 sequences. Among particularly desirable AAV-1 fragments are the inverted terminal repeat sequences (ITRs), rep and cap. Each of these fragments may be readily utilized, e.g., as a cassette, in a variety of vector systems and host cells. Such fragments may be used alone, in combination with other AAV-1 sequences or fragments, or in combination with elements from other AAV or non-AAV viral sequences. In one particularly desirable embodiment, a cassette may contain the AAV-1 ITRs of the invention flanking a selected transgene. In another desirable embodiment, a cassette may contain the AAV-1 rep and/or cap proteins, e.g., for use in producing recombinant (rAAV) virus.

Thus, the AAV-1 sequences and fragments thereof are useful in production of rAAV, and are also useful as antisense delivery vectors, gene therapy vectors, or vaccine vectors. The invention further provides nucleic acid molecules, gene delivery

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vectors, and host cells which contain the AAV-1 sequences of the invention. Also provided a novel methods of gene delivery using AAV vectors.

As described herein, the vectors of the invention containing the AAV-1 capsid proteins of the invention are particularly well suited for use in applications in which the neutralizing antibodies diminish the effectiveness of other AAV serotype based vectors, as well as other viral vectors. The rAAV vectors of the invention are particularly advantageous in rAAV readministration and repeat gene therapy.

These and other embodiments and advantages of the invention are described in more detail below. As used throughout this specification and the claims, the term "comprising" is inclusive of other components, elements, integers, steps and the like.

### I. AAV-1 NUCLEIC ACID AND PROTEIN SEQUENCES

The AAV-1 nucleic acid sequences of the invention include the DNA sequences of SEQ ID NO: 1 (Figs. 1A-1C), which consists of 4718 nucleotides. The AAV-1 nucleic acid sequences of the invention further encompass the strand which is complementary to SEQ ID NO: 1, as well as the RNA and cDNA sequences corresponding to SEQ ID NO: 1 and its complementary strand. Also included in the nucleic acid sequences of the invention are natural variants and engineered modifications of SEQ ID NO: 1 and its complementary strand. Such modifications include, for example, labels which are known in the art, methylation, and substitution of one or more of the naturally occurring nucleotides with an analog.

Further included in this invention are nucleic acid sequences which are greater than 85%, preferably at least about 90%, more preferably at least about 95%, and most preferably at least about 98 - 99% identical or homologous to SEQ ID NO:1. The term "percent sequence identity" or "identical" in the context of nucleic acid sequences refers to the residues in the two sequences which are the same when aligned for maximum correspondence. The length of sequence identity comparison may be over the full-length sequence, or a fragment at least about nine nucleotides, usually at least about 20 - 24 nucleotides, at least about 28 - 32 nucleotides, and preferably at least about 36 or more nucleotides. There are a number of different

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algorithms known in the art which can be used to measure nucleotide sequence identity. For instance, polynucleotide sequences can be compared using Fasta, a program in GCG Version 6.1. Fasta provides alignments and percent sequence identity of the regions of the best overlap between the query and search sequences (Pearson, 1990, herein incorporated by reference). For instance, percent sequence identity between nucleic acid sequences can be determined using Fasta with its default parameters (a word size of 6 and the NOPAM factor for the scoring matrix) as provided in GCG Version 6.1, herein incorporated by reference.

The term "substantial homology" or "substantial similarity," when referring to a nucleic acid or fragment thereof, indicates that, when optimally aligned with appropriate nucleotide insertions or deletions with another nucleic acid (or its complementary strand), there is nucleotide sequence identity in at least about 95 - 99% of the sequence.

Also included within the invention are fragments of SEQ ID NO: 1, its complementary strand, cDNA and RNA complementary thereto. Suitable fragments are at least 15 nucleotides in length, and encompass functional fragments which are of biological interest. Certain of these fragments may be identified by reference to Figs. 1A-1C. Examples of particularly desirable functional fragments include the AAV-1 inverted terminal repeat (ITR) sequences of the invention. In contrast to the 145 nt ITRs of AAV-2, AAV-3, and AAV-4, the AAV-1 ITRs have been found to consist of only 143 nucleotides, yet advantageously are characterized by the T-shaped hairpin structure which is believed to be responsible for the ability of the AAV-2 ITRs to direct site-specific integration. In addition, AAV-1 is unique among other AAV serotypes, in that the 5' and 3' ITRs are identical. The full-length 5' ITR sequences of AAV-1 are provided at nucleotides 1-143 of SEQ ID NO: 1 (Fig. 1A) and the fulllength 3' ITR sequences of AAV-1 are provided at nt 4576-4718 of SEQ ID NO: 1 (Fig. 1C). One of skill in the art can readily utilize less than the full-length 5' and/or 3' ITR sequences for various purposes and may construct modified ITRs using conventional techniques, e.g., as described for AAV-2 ITRs in Samulski et al, Cell, <u>33</u>:135-143 (1983).

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Another desirable functional fragment of the AAV-1 genome is the P5 promoter of AAV-1 which has sequences unique among AAV P5 promoters, while maintaining critical regulatory elements and functions. This promoter is located within nt 236 - 299 of SEQ ID NO: 1 (Fig. 1A). Other examples of functional fragments of interest include the sequences at the junction of the rep/cap, e.g., the sequences spanning nt 2306-2223, as well as larger fragments which encompass this junction which may comprise 50 nucleotides on either side of this junction. Still other examples of functional fragments include the sequences encoding the rep proteins. Rep 78 is located in the region of nt 334 - 2306 of SEQ ID NO: 1; Rep 68 is located in the region of nt 334-2272, and contains an intron spanning nt 1924-2220 of SEQ ID NO: 1. Rep 52 is located in the region of nt 1007 - 2304 of SEQ ID NO: 1; rep 40 is located in the region of nt 1007 - 2272, and contains an intron spanning nt 1924-2246 of SEO ID NO: 1. Also of interest are the sequences encoding the capsid proteins, VP 1 [nt 2223-4431 of SEQ ID NO: 1], VP2 [nt 2634-4432 of SEQ ID NO: 1] and VP3 [nt 2829-4432 of SEQ ID NO: 1]. Other fragments of interest may include the AAV-1 P19 sequences, AAV-1 P40 sequences, the rep binding site, and the terminal resolute site (TRS).

The invention further provides the proteins and fragments thereof which are encoded by the AAV-1 nucleic acids of the invention. Particularly desirable proteins include the rep and cap proteins, which are encoded by the nucleotide sequences identified above. These proteins include rep 78 [SEQ ID NO:5], rep 68 [SEQ ID NO:7], rep 52 [SEQ ID NO:9], rep 40 [SEQ ID NO: 11], vpl [SEQ ID NO: 13], vp2 [SEQ ID NO: 15], and vp3 [SEQ IID NO: 17] and functional fragments thereof while the sequences of the rep and cap proteins have been found to be closely related to those of AAV-6, there are differences in the amino acid sequences (see Table 1 below), as well as differences in the recognition of these proteins by the immune system. However, one of skill in the art may readily select other suitable proteins or protein fragments of biological interest. Suitably, such fragments are at least 8 amino acids in length. However, fragments of other desired lengths may be readily utilized.

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Such fragments may be produced recombinantly or by other suitable means, e.g., chemical synthesis.

The sequences, proteins, and fragments of the invention may be produced by any suitable means, including recombinant production, chemical synthesis, or other synthetic means. Such production methods are within the knowledge of those of skill in the art and are not a limitation of the present invention.

### II. VIRAL VECTORS

In another aspect, the present invention provides vectors which utilize the AAV-1 sequences of the invention, including fragments thereof, for delivery of a heterologous gene or other nucleic acid sequences to a target cell. Suitably, these heterologous sequences (i.e., a transgene) encode a protein or gene product which is capable of being expressed in the target cell. Such a transgene may be constructed in the form of a "minigene". Such a "minigene" includes selected heterologous gene sequences and the other regulatory elements necessary to transcribe the gene and express the gene product in a host cell. Thus, the gene sequences are operatively linked to regulatory components in a manner which permit their transcription. Such components include conventional regulatory elements necessary to drive expression of the transgene in a cell containing the viral vector. The minigene may also contain a selected promoter which is linked to the transgene and located, with other regulatory elements, within the selected viral sequences of the recombinant vector.

Selection of the promoter is a routine matter and is not a limitation of this invention. Useful promoters may be constitutive promoters or regulated (inducible) promoters, which will enable control of the timing and amount of the transgene to be expressed. For example, desirable promoters include the cytomegalovirus (CMV) immediate early promoter/enhancer [see, e.g., Boshart et al, Cell, 41:521-530 (1985)], the Rous sarcoma virus LTR promoter/enhancer, and the chicken cytoplasmic β-actin promoter [T. A. Kost et al, Nucl, Acids Res., 11(23):8287 (1983)]. Still other desirable promoters are the albumin promoter and an AAV P5 promoter. Optionally, the selected promoter is used in conjunction with a heterologous enhancer, e.g., the β-

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actin promoter may be used in conjunction with the CMV enhancer. Yet other suitable or desirable promoters and enhancers may be selected by one of skill in the art.

The minigene may also desirably contain nucleic acid sequences heterologous to the viral vector sequences including sequences providing signals required for efficient polyadenylation of the transcript (poly-A or pA) and introns with functional splice donor and acceptor sites. A common poly-A sequence which is employed in the exemplary vectors of this invention is that derived from the papovavirus SV-40. The poly-A sequence generally is inserted in the minigene downstream of the transgene sequences and upstream of the viral vector sequences. A common intron sequence is also derived from SV-40, and is referred to as the SV40 T intron sequence. A minigene of the present invention may also contain such an intron, desirably located between the promoter/enhancer sequence and the transgene. Selection of these and other common vector elements are conventional [see, e.g., Sambrook et al, "Molecular Cloning. A Laboratory Manual", 2d edit., Cold Spring Harbor Laboratory, New York (1989) and references cited therein] and many such sequences are available from commercial and industrial sources as well as from Genebank.

The selection of the transgene is not a limitation of the present invention. Suitable transgenes may be readily selected from among desirable reporter genes, therapeutic genes, and optionally, genes encoding immunogenic polypeptides. Examples of suitable reporter genes include β-galactosidase (β-gal), an alkaline phosphatase gene, and green fluorescent protein (GFP). Examples of therapeutic genes include, cytokines, growth factors, hormones, and differentiation factors, among others. The transgene may be readily selected by one of skill in the art. See, e.g., WO 98/09657, which identifies other suitable transgenes.

Suitably, the vectors of the invention contain, at a minimum, cassettes which consist of fragments of the AAV-1 sequences and proteins. In one embodiment, a vector of the invention comprises a selected transgene, which is flanked by a 5' ITR and a 3' ITR, at least one of which is an AAV-1 ITR of the invention. Suitably,

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vectors of the invention may contain a AAV-1 P5 promoter of the invention. In yet another embodiment, a plasmid or vector of the invention contains AAV-1 rep sequences. In still another embodiment, a plasmid or vector of the invention contains at least one of the AAV-1 cap proteins of the invention. Most suitably, these AAV-1-derived vectors are assembled into viral vectors, as described herein.

#### A. AAV Viral Vectors

In one aspect, the present invention provides a recombinant AAV-1 viral vector produced using the AAV-1 capsid proteins of the invention. The packaged rAAV-1 virions of the invention may contain, in addition to a selected minigene, other AAV-1 sequences, or may contain sequences from other AAV serotypes.

Methods of generating rAAV virions are well known and the selection of a suitable method is not a limitation on the present invention. See, e.g., K. Fisher et al, J. Virol., 70:520-532 (1993) and US Patent 5,478,745. In one suitable method, a selected host cell is provided with the AAV sequence encoding a rep protein, the gene encoding the AAV cap protein and with the sequences for packaging and subsequent delivery. Desirably, the method utilizes the sequences encoding the AAV-1 rep and/or cap proteins of the invention.

In one embodiment, the rep/cap genes and the sequences for delivery are supplied by co-transfection of vectors carrying these genes and sequences. In one currently preferred embodiment, a cis (vector) plasmid, a trans plasmid containing the rep and cap genes, and a plasmid containing the adenovirus helper genes are co-transfected into a suitable cell line, e.g., 293. Alternatively, one or more of these functions may be provided in trans via separate vectors, or may be found in a suitably engineered packaging cell line.

An exemplary cis plasmid will contain, in 5' to 3' order, AAV 5' ITR, the selected transgene, and AAV 3' ITR. In one desirable embodiment, at least one of the AAV ITRs is a 143 nt AAV-1 ITR. However, other AAV serotype ITRs may be readily selected. Suitably, the full-length ITRs are utilized. However, one of skill in

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the art can readily prepare modified AAV ITRs using conventional techniques.

Similarly, methods for construction of such plasmids is well known to those of skill in the art.

A trans plasmid for use in the production of the rAAV-1 virion particle may be prepared according to known techniques. In one desired embodiment, this plasmid contains the rep and cap proteins of AAV-1, or functional fragments thereof. Alternatively, the rep sequences may be from another selected AAV serotype.

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The cis and trans plasmid may then be co-transfected with a wild-type helper virus (e.g., Ad2, Ad5, or a herpesvirus), or more desirably, a replication - defective adenovirus, into a selected host cell. Alternatively, the cis and trans plasmid may be co-transfected into a selected host cell together with a transfected plasmid which provides the necessary helper functions. Selection of a suitable host cell is well within the skill of those in the art and include such mammalian cells as 293 cells, HeLa cells, among others.

Alternatively, the cis plasmid and, optionally the trans plasmid, may be transfected into a packaging cell line which provides the remaining helper functions necessary for production of a rAAV containing the desired AAV-1 sequences of the invention. An example of a suitable packaging cell line, where an AAV-2 capsid is desired, is B-50, which stably expresses AAV-2 rep and cap genes under the control of a homologous P5 promoter. This cell line is characterized by integration into the cellular chromosome of multiple copies (at least 5 copies) of P5-rep-cap gene cassettes in a concatomer form. This B-50 cell line was deposited with the American Type Culture Collection, 10801 University Boulevard, Manassas, Virginia 20110-2209, on September 18, 1997 under Accession No. CRL-12401 pursuant to the provisions of the Budapest Treaty. However, the present invention is not limited as to the selection of the packaging cell line.

Exemplary transducing vectors based on AAV-1 capsid proteins have been tested both *in vivo and in vitro*, as described in more detail in Example 4. In these studies, it was demonstrated that recombinant AAV vector with an AAV-1 virion can transduce both mouse liver and muscle. These, and other AAV-1 based

gene therapy vectors which may be generated by one of skill in the art are beneficial for gene delivery to selected host cells and gene therapy patients since the neutralization antibodies of AAV-1 present in much of the human population exhibit different patterns from other AAV serotypes and therefore do not neutralize the AAV-1 virions. One of skill in the art may readily prepare other rAAV viral vectors containing the AAV-1 capsid proteins provided herein using a variety of techniques known to those of skill in the art. One may similarly prepare still other rAAV viral vectors containing AAV-1 sequence and AAV capsids of another serotype.

#### B. Other Viral Vectors

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One of skill in the art will readily understand that the AAV-1 sequences of the invention can be readily adapted for use in these and other viral vector systems for *in vitro*, *ex vivo or in vivo* gene delivery. Particularly well suited for use in such viral vector systems are the AAV-1 ITR sequences, the AAV-1 rep, the AAV-1 cap, and the AAV-1 P5 promoter sequences.

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For example, in one desirable embodiment, the AAV-1 ITR sequences of the invention may be used in an expression cassette which includes AAV-1 5' ITR, a non-AAV DNA sequences of interest (e.g., a minigene), and 3' ITR and which lacks functional rep/cap. Such a cassette containing an AAV-1 ITR may be located on a plasmid for subsequent transfection into a desired host cell, such as the cis plasmid described above. This expression cassette may further be provided with an AAV capsid of a selected serotype to permit infection of a cell or stably transfected into a desired host cell for packaging of rAAV virions. Such an expression cassette may be readily adapted for use in other viral systems, including adenovirus systems and lentivirus systems. Methods of producing Ad/AAV vectors are well known to those of skill in the art. One desirable method is described in PCT/US95/14018. However, the present invention is not limited to any particular method.

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Another aspect of the present invention is the novel AAV-1 P5 promoter sequences which are located in the region spanning nt 236 - 299 of SEQ ID NO: 1. This promoter is useful in a variety of viral vectors for driving expression of a desired transgene.

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Similarly, one of skill in the art can readily select other fragments of the AAV-1 genome of the invention for use in a variety of vector systems. Such vectors systems may include, e.g., lentiviruses, retroviruses, poxviruses, vaccinia viruses, and adenoviral systems, among others. Selection of these vector systems is not a limitation of the present invention.

### C. Host Cells And Packaging Cell Lines

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In yet another aspect, the present invention provides host cells which may be transiently transfected with AAV-1 nucleic acid sequences of the invention to permit expression of a desired transgene or production of a rAAV particle. For example, a selected host cell may be transfected with the AAV-1 P5 promoter sequences and/or the AAV-1 5' ITR sequences using conventional techniques. Providing AAV helper functions to the transfected cell lines of the invention results in packaging of the rAAV as infectious rAAV particles. Such cell lines may be produced in accordance with known techniques [see, e.g, US Patent No. 5,658,785], making use of the AAV-1 sequences of the invention.

Alternatively, host cells of the invention may be stably transfected with a rAAV expression cassette of the invention, and with copies of AAV-1 rep and cap genes. Suitable parental cell lines include mammalian cell lines and it may be desirable to select host cells from among non-simian mammalian cells. Examples of suitable parental cell lines include, without limitation, HeLa [ATCC CCL 2], A549 [ATCC Accession No. CCL 185], KB [CCL 17], Detroit [e.g., Detroit 510, CCL 72] and WI-38 [CCL 75] cells. These cell lines are all available from the American Type Culture Collection, 10801 University Boulevard, Manassas, Virginia 20110-2209 USA. Other suitable parent cell lines may be obtained from other sources and may be used to construct stable cell lines containing the P5 and/or AAV rep and cap sequences of the invention.

Recombinant vectors generated as described above are useful for delivery of the DNA of interest to cells.

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### III. METHODS OF DELIVERING GENES VIA AAV-1 DERIVED VECTORS

In another aspect, the present invention provides a method for delivery of a transgene to a host which involves transfecting or infecting a selected host cell with a recombinant viral vector generated with the AAV-1 sequences (or functional fragments thereof) of the invention. Methods for delivery are well known to those of skill in the art and are not a limitation of the present invention.

In one desirable embodiment, the invention provides a method for AAV-mediated delivery of a transgene to a host. This method involves transfecting or
infecting a selected host cell with a recombinant viral vector containing a selected
transgene under the control of sequences which direct expression thereof and AAV-1
capsid proteins.

Optionally, a sample from the host may be first assayed for the presence of antibodies to a selected AAV serotype. A variety of assay formats for detecting neutralizing antibodies are well known to those of skill in the art. The selection of such an assay is not a limitation of the present invention. See, e.g., Fisher et al, Nature Med., 3(3):306-312 (March 1997) and W. C. Manning et al, Human Gene Therapy, 9:477-485 (March 1, 1998). The results of this assay may be used to determine which AAV vector containing capsid proteins of a particular serotype are preferred for delivery, e.g., by the absence of neutralizing antibodies specific for that capsid serotype.

In one aspect of this method, the delivery of vector with AAV-1 capsid proteins may precede or follow delivery of a gene via a vector with a different serotype AAV capsid protein. Thus, gene delivery via rAAV vectors may be used for repeat gene delivery to a selected host cell. Desirably, subsequently administered rAAV vectors carry the same transgene as the first rAAV vector, but the subsequently administered vectors contain capsid proteins of serotypes which differ from the first vector. For example, if a first vector has AAV-2 capsid proteins, subsequently administered vectors may have capsid proteins selected from among the other serotypes, including AAV-1, AAV-3A, AAV-3B, AAV-4 and AAV-6.

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Thus, a rAAV-1-derived recombinant viral vector of the invention provides an efficient gene transfer vehicle which can deliver a selected transgene to a selected host cell *in vivo or ex vivo* even where the organism has neutralizing antibodies to one or more AAV serotypes. These compositions are particularly well suited to gene delivery for therapeutic purposes. However, the compositions of the invention may also be useful in immunization. Further, the compositions of the invention may also be used for production of a desired gene product *in vitro*.

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The above-described recombinant vectors may be delivered to host cells according to published methods. An AAV viral vector bearing the selected transgene may be administered to a patient, preferably suspended in a biologically compatible solution or pharmaceutically acceptable delivery vehicle. A suitable vehicle includes sterile saline. Other aqueous and non-aqueous isotonic sterile injection solutions and aqueous and non-aqueous sterile suspensions known to be pharmaceutically acceptable carriers and well known to those of skill in the art may be employed for this purpose.

The viral vectors are administered in sufficient amounts to transfect the cells and to provide sufficient levels of gene transfer and expression to provide a therapeutic benefit without undue adverse effects, or with medically acceptable physiological effects, which can be determined by those skilled in the medical arts. Conventional and pharmaceutically acceptable routes of administration include, but are not limited to, direct delivery to the liver, oral, intranasal, intravenous, intramuscular, subcutaneous, intradermal, and other parental routes of administration. Routes of administration may be combined, if desired.

Dosages of the viral vector will depend primarily on factors such as the condition being treated, the age, weight and health of the patient, and may thus vary among patients. For example, a therapeutically effective human dosage of the viral vector is generally in the range of from about 1 ml to about 100 ml of solution containing concentrations of from about 1 x 10<sup>9</sup> to 1 x 10<sup>16</sup> genomes virus vector. A preferred human dosage may be about 1 x 10<sup>13</sup> to 1 x 10<sup>16</sup> AAV genomes. The dosage will be adjusted to balance the therapeutic benefit against any side effects and

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such dosages may vary depending upon the therapeutic application for which the recombinant vector is employed. The levels of expression of the transgene can be monitored to determine the frequency of dosage resulting in viral vectors, preferably AAV vectors containing the minigene. Optionally, dosage regimens similar to those described for therapeutic purposes may be utilized for immunization using the compositions of the invention. For *in vitro* production, a desired protein may be obtained from a desired culture following transfection of host cells with a rAAV containing the gene encoding the desired protein and culturing the cell culture under conditions which permits expression. The expressed protein may then be purified and isolated, as desired. Suitable techniques for transfection, cell culturing, purification, and isolation are known to those of skill in the art.

The following examples illustrate several aspects and embodiments of the invention.

### Example 1 - Generation of Infectious Clone of AAV-1

The replicated form DNA of AAV-1 was extracted from 293 cells that were infected by AAV-1 and wild type adenovirus type 5.

### A. Cell Culture and Virus

AAV-free 293 cells and 84-31 cells were provided by the human application laboratory of the University of Pennsylvania. These cells were cultured in Dulbecco's Modified Eagle Medium with 10% fetal bovine serum (Hyclone), penicillin (100 U/ml) and streptomycin at 37°C in a moisturized environment supplied with 5% C0<sub>2</sub>. The 84-31 cell line constitutively expresses adenovirus genes E1a, Elb, E4/ORF6, and has been described previously [K. J. Fisher, J. Virol., 70:520-532 (1996)]. AAV-1 (ATCC VR-645) seed stock was purchased from American Type Culture Collection (ATCC, Manassas, VA). AAV viruses were propagated in 293 cells with wild type Ad5 as a helper virus.

### B. Recombinant AAV Generation

The recombinant AAV viruses were generated by transfection using an adenovirus free method. Briefly, the cis plasmid (with AAV ITR), trans plasmid (with

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AAV rep gene and cap gene) and helper plasmid (pFa13, with essential regions from the adenovirus genome) were simultaneously co-transfected into 293 cells in a ratio of 1:1:2 by calcium phosphate precipitation. The pFa13 helper plasmid has an 8 kb deletion in the adenovirus E2B region and has deletions in most of the late genes. This helper plasmid was generated by deleting the RsrII fragment from pFG140 (Microbix, Canada). Typically, 50 µg of DNA (cis:trans:PFa13 at ratios of 1:1:2, respectively) was transfected onto a 15 cm tissue culture dish. The cells were harvested 96 hours post-transfection, sonicated and treated with 0.5% sodium deoxycholate (37°C for 10 min). Cell lysates were then subjected to two rounds of a CsCl gradient. Peak fractions containing AAV vector were collected, pooled, and dialyzed against PBS before injecting into animals. To make rAAV virus with AAV-1 virion, the pAV1H or p5E18 (2/1) was used as the *trans* plasmid to provide rep and cap function.

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For the generation of rAAV based on AAV-2, p5E18 was used as the trans plasmid since it greatly improved the rAAV yield. This plasmid, p5E18(2/2), expresses AAV-2 Rep and Cap and contains a P5 promoter relocated to a position 3' to the Cap gene, thereby minimizing expression of Rep78 and Rep68. The strategy was initially described by Li et al, <u>J. Virol.</u>, <u>71</u>:5236-5243 (1997). P5E18(2/2) was constructed in the following way. The previously described pMMTV-trans vector (i.e., the mouse mammary tumor virus promoter substituted for the P5 promoter in an AAV-2-based vector) was digested with Smal and ClaI, filled in with the Klenow enzyme, and then recircularized with DNA ligase. The resulting construct was digested with XbaI, filled in, and ligated to the blunt-ended BamHI-XbaI fragment from pCR-p5, constructed in the following way. The P5 promoter of AAV was amplified by PCR and the amplified fragment was subsequently cloned into pCR2.1 (Invitrogen) to yield pCR-P5. The helper plasmid pAV1H was constructed by cloning the BfaI fragment of pAAV-2 into pBluescript II-SK(+) at the BcorV and SmaI sites. The 3.0-kb XbaI-KpnI fragment from p5E18(2/2), the 2.3-kb XbaI-KpnI fragment from pAV1H, and the 1.7-kb KpnI fragment from p5E18(2/2) were incorporated into a separate plasmid P5E18(2/1), which contains AAV-2 Rep, AAV-1 Cap, and the

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AAV-2 P5 promoter located 3' to the Cap gene. Plasmid p5E18(2/1) produced 10- to 20-fold higher quantities of the vector than pAV1H (i.e., 10<sup>12</sup> genomes/50 15-cm<sup>2</sup> plates).

### C. <u>DNA Techniques</u>

5 Hirt DNA extraction was performed as described in the art with minor modification [R.J. Samulski et al., Cell, 33:135-143 (1983)]. More particularly, Hirst solution without SDS was used instead of using original Hirt solution containing SDS. The amount of SDS present in the original Hirst solution was added after the cells had been fully suspended. To construct AAV-1 infectious clone, the Hirt DNA from AAV-1 infected 293 cells was repaired with Klenow enzyme (New England Biolabs) 10 to ensure the ends were blunt. The treated AAV-1 Hirt DNA was then digested with BamHI and cloned into three vectors, respectively. The internal BamHI was cloned into pBlueScript II-SK+ cut with BamHI to get pAVI-BM. The left and right fragments were cloned into pBlueScript II-SK+ cut with BamHI + EcoRV to obtain 15 pAV1-BL and pAV1-BR, respectively. The AAV sequence in these three plasmids were subsequently assembled into the same vector to get AAV-1 infectious clone pAAV-1. The helper plasmid for recombinant AAV-1 virus generation was constructed by cloning the Bfa I fragment of pAAV-1 into pBlueScript II-SK+ at the EcoRV site.

Analysis of the Hirt DNA revealed three bands, a dimer at 9.4 kb, a monomer at 4.7 kb and single-stranded DNA at 1.7 kb, which correlated to different replication forms of AAV-1. The monomer band was excised from the gel and then digested with *BamH*I. This resulted in three fragments of 1.1 kb, 0.8 kb and 2.8 kb. This pattern is in accordance with the description by Bantel-schaal and zur Hausen, Virol., 134(1):52-63 (1984). The 1.1 kb and 2.8 kb *BamH*I fragments were cloned into pBlueScript-KS(+) at *BamH*I and EcoRV site. The internal 0.8 kb fragment was cloned into *BamH*I site of pBlueScript-KS(+).

These three fragments were then subcloned into the same construct to obtain a plasmid (pAAV-1) that contained the full sequence of AAV-1. The pAAV-1 was then tested for its ability to rescue from the plasmid backbone and package

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infectious virus. The pAAV-1 was then transfected to 293 cells and supplied with adenovirus type as helper at MOI 10. The virus supernatant was used to reinfect 293 cells.

For Southern blot analysis, Hirt DNA was digested with *Dpn*I to remove bacteria-borne plasmid and probed with internal *BamH*I fragment of AAV-1. The membrane was then washed at high stringency conditions, which included: twice 30 minutes with 2X SSC, 0.1% SDS at 65°C and twice 30 minutes with 0.1X SSC, 0.1% SDS at 65°C. The membrane was then analyzed by both phosphor image and X-ray autoradiography. The results confirmed that pAAV-1 is indeed an infectious clone of AAV serotype 1.

### Example 2 - Sequencing Analysis of AAV-1

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The entire AAV-1 genome was then determined by automatic sequencing and was found to be 4718 nucleotides in length (Figs. 1A-1C). For sequencing, an ABI 373 automatic sequencer as used to determine the sequences for all plasmids and PCR fragments related to this study using the FS dye chemistry. All sequences were confirmed by sequencing both plus and minus strands. These sequences were also confirmed by sequencing two independent clones of pAV-BM, pAV-BL and pAV-BR. Since the replicated form of AAV-1 DNA served as the template for sequence determination, these sequences were also confirmed by sequencing a series of PCR products using original AAV-1 seed stock as a template.

The length of AAV-1 was found to be within the range of the other serotypes: AAV-3 (4726 nucleotides), AAV-4 (4774 nucleotides), AAV-2 (4681 nucleotides), and AAV-6 (4683 nucleotides).

The AAV-1 genome exhibited similarities to other serotypes of adenoassociated viruses. Overall, it shares more than 80% identity with other known AAV
viruses as determined by the computer program Megalign using default settings
[DNASTAR, Madison, WI]. The key features in AAV-2 can also be found in AAV1. First, AAV-1 has the same type of inverted terminal repeat which is capable of
forming T-shaped hairpin structures, despite the differences at the nucleotide level

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(Figs. 2 and 3). The sequences of right ITRs and left ITRs of AAV-1 are identical. The AAV TR sequence is subdivided into A, A', B, B', C, C', D and D' [Bern, cited above].

These AAV ITR sequences are also virtually the same as those found in AAV-6 right ITR, there being one nucleotide difference in each of A and A' sequence, and the last nucleotide of the D sequence. Second, the AAV-2 rep binding motif [GCTCGCTCGCTCGCTG (SEQ ID NO: 20)] is well conserved. Such motif can also be found in the human chromosome 19 AAV-2 pre-integration region. Finally, non-structural and structural coding regions, and regulatory elements similar to those of other AAV serotypes also exist in AAV-1 genome.

Although the overall features of AAV terminal repeats are very much conserved, the total length of the AAV terminal repeat exhibits divergence. The terminal repeat of AAV-1 consists of 143 nucleotides while those of AAV-2, AAV-3, and AAV-4 are about 145 or 146 nucleotides. The loop region of AAV-1 ITR most closely resembles that of AAV-4 in that it also uses TCT instead of the TTT found in AAV-2 and AAV-3. The possibility of sequencing error was eliminated using restriction enzyme digestion, since these three nucleotides are part of the SacI site (gagete; nt 69-74 of SEQ ID NO: 1). The p5 promoter region of AAV-1 shows more variations in nucleotide sequences with other AAV serotypes. However, it still maintains the critical regulatory elements. The two copies of YY1 [See, Fig. 1A-1C] sites seemed to be preserved in all known AAV serotypes, which have been shown to be involved in regulating AAV gene expression. In AAV-4, there are 56 additional nucleotides inserted between YY1 and E-box/USF site, while in AAV-1, there are 26 additional nucleotides inserted before the E-box/USF site. The p19 promoter, p40 promoter and polyA can also be identified from the AAV-1 genome by analogy to known AAV serotypes, which are also highly conserved.

Thus, the analysis of AAV terminal repeats of various serotypes showed that the A and A' sequence is very much conserved. One of the reasons may be the Rep binding motif (GCTC)<sub>3</sub>GCTG [SEQ ID NO: 20]. These sequences appear to be essential for AAV DNA replication and site-specific integration. The same sequence

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has also been shown to be preserved in a monkey genome [Samulski, personal communication]. The first 8 nucleotides of the D sequence are also identical in all known AAV serotypes. This is in accordance with the observation of the Srivastava group that only the first 10 nucleotides are essential for AAV packaging [X.S. Wang et al, J. Virol., 71:3077-3082 (1997); X.S. Wang et al, J. Virol., 71:1140-1146 (1997)]. The function of the rest of the D sequences still remain unclear. They may be somehow related to their tissue specificities. The variation of nucleotide in B and C sequence may also suggest that the secondary structure of the ITRs is more critical for its biological function, which has been demonstrated in many previous publications.

### Example 3 - Comparison of AAV-1 Sequences

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The nucleotide sequences of AAV-1, obtained as described above, were compared with known AAV sequences, including AAV-2, AAV-4 and AAV-6 using DNA Star Megalign. This comparison revealed a stretch of 71 identical nucleotides shared by AAV-1, AAV-2 and AAV-6. See, Figs. 1A-1C.

This comparison further suggested that AAV-6 is a hybrid formed by homologous recombination of AAV-1 and AAV-2. See, Figs. 3A and 3B. These nucleotides divide the AAV-6 genome into two regions. The 5' half of AAV-6 of 522 nucleotides is identical to that of AAV-2 except in 2 positions. The 3' half of AAV-6 including the majority of the rep gene, complete cap gene and 3' ITR is 98% identical to AAV-1.

Biologically, such recombination may enable AAV-1 to acquire the ability to transmit through the human population. It is also interesting to note that the ITRs of AAV-6 comprise one AAV-1 ITR and one AAV-2 ITR. The replication model of defective parvovirus can maintain this special arrangement. Studies on AAV integration have shown that a majority of AAV integrants carries deletions in at least one of the terminal repeats. These deletions have been shown to be able to be repaired through gene conversion using the other intact terminal repeat as a template. Therefore, it would be very difficult to maintain AAV-6 as a homogenous population

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when an integrated copy of AAV-6 is rescued from host cells with helper virus infection. The AAV-6 with two identical AAV-2 ITRs or two identical AAV-1 ITRs should be the dominant variants. The AAV-6 with two AAV-1 ITRs has been observed by Russell's group [Rutledge, cited above (1998)]. So far there is no report on AAV-6 with two AAV-2 ITRs. Acquirement of AAV-2 P5 promoter by AAV-6 may have explained that AAV-6 have been isolated from human origin while AAV-1 with the same virion has not. The regulation of P5 promoter between different species of AAV may be different *in vivo*. This observation suggests the capsid proteins of AAV were not the only determinants for tissue specificity.

Although it is clear that AAV-6 is a hybrid of AAV-1 and AAV-2, AAV-6 has already exhibited divergence from either AAV-1 or AAV-2. There are two nucleotide differences between AAV-6 and AAV-2 in their first 450 nucleotides. There are about 1% differences between AAV-6 and AAV-1 in nucleotide levels from nucleotides 522 to the 3' end. There also exists a quite divergent region (nucleotide 4486-4593) between AAV-6 and AAV-1 (Figs. 1A-1C). This region does not encode any known proteins for AAVs. These differences in nucleotide sequences may suggest that AAV-6 and AAV-1 have gone through some evolution since the recombination took place. Another possible explanation is that there exists another variant of AAV-1 which has yet to be identified. So far, there is no evidence to rule out either possibility. It is still unknown if other hybrids (AAV-2 to AAV-4, etc.) existed in nature.

The coding region of AAV-1 was deduced by comparison with other known AAV serotypes. Table 1 illustrates the coding region differences between AAV-1 and AAV-6. The amino acid residues are deduced according to AAV-2.

With reference to the amino acid position of AAV-1, Table 1 lists the amino acids of AAV-1 which have been changed to the corresponding ones of AAV-6. The amino acids of AAV-1 are shown to the left of the arrow. Reference may be made to SEQ ID NO: 5 of the amino acid sequence of AAV-1 Rep 78 and to SEQ ID NO: 13 for the amino acid sequence of AAV-1 VP1.

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Table 1

Coding region variations between AAV-1 and AAV-6

Rep prote	ein (Rep78)		Cap pro	tein (VP1)
Position(s)	sition(s) Amino acids		Position(s)	Amino acids
28	S-N		129	L→F
191	Q−H		418	E-D
192	H-D		531	E→K
308	E-D	1	584	F-L
			598	A-V
:			642	N-H

It was surprising to see that the sequence of the AAV-1 coding region is almost identical to that of AAV-6 from position 452 to the end of coding region (99%). The first 508 nucleotides of AAV-6 have been shown to be identical to those of AAV-2 [Rutledge, cited above (1998)]. Since the components of AAV-6 genome seemed to be AAV-2 left ITR - AAV-2 p5 promoter - AAV-1 coding region - AAV-1 right ITR, it was concluded that AAV-6 is a naturally occurred hybrid between AAV-1 and AAV-2.

### Example 4 - Gene Therapy Vector Based on AAV-1

Recombinant gene transfer vectors based on AAV-1 viruses were constructed by the methods described in Example 1. To produce a hybrid recombinant virus with AAV-1 virion and AAV-2 ITR, the AAV-1 trans plasmid (pAV1H) and the AAV-2 cis-lacZ plasmid (with AAV-2 ITR) were used. The AAV-2 ITR was used in this vector in view of its known ability to direct site-specific integration. Also constructed for use in this experiment was an AAV-1 vector carrying the green fluorescent protein (GFP) marker gene under the control of the immediate early promoter of CMV using pAV1H as the trans plasmid.

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### A. rAAV-1 Viruses Transfect Host Cells in Vitro

84-31 cells, which are subclones of 293 cells (which express adenovirus E1a, E1b) which stably express E4/ORF5, were infected with rAAV-1 GFP or rAAV-lacZ. High levels of expression of GFP and lacZ was detected in the cultured 84-31 cells. This suggested that rAAV-1 based vector was very similar to AAV-2 based vectors in ability to infect and expression levels.

### B. rAAV-1 Viruses Transfect Cells in Vivo

The performance of AAV-1 based vectors was also tested *in vivo*. The rAAV-1 CMV-α1AT virus was constructed as follows. The EcoRI fragment of pAT85 (ATCC) containing human α1-antitrypsin (α1AT) cDNA fragment was blunted and cloned into PCR (Promega) at a Smal site to obtain PCR-α1AT. The CMV promoter was cloned into PCR-α1AT at the XbaI site. The Alb-α1AT expression cassette was removed by Xhol and ClaI and cloned into pAV1H at the XbaI site. This vector plasmid was used to generate AAV-1-CMV-α1AT virus used in the experiment described below.

For screening human antibodies against AAV, purified AAV virus is lysed with Ripa buffer (10 mM Tris pH 8.2, 1% Triton X-100, 1% SDS, 0.15 M NaCl) and separated in 10% SDS-PAGE gel. The heat inactivated human serum was used at a 1 to 1000 dilution in this assay. The rAAV-1 CMV-α1AT viruses were injected into Rag-1 mice through tail vein injection at different dosages. The concentration of human α1-antitrypsin in mouse serum was measured using ELISA. The coating antibody is rabbit anti-human human α1-antitrypsin (Sigma). The goat-antihuman α1-antitrypsin (Sigma) was used as the primary detection antibodies. The sensitivity of this assay is around 0.3 ng/ml to 30 ng/ml. The expression of human α-antitrypsin in mouse blood can be detected in a very encouraging level. This result is shown in Table 2.

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Table 2
Human Antitrypsin Expressed in Mouse Liver

Amount of virus injected	Week 2 (ng/ml)	Week 4 (ng/ml)	
2x10 <sup>10</sup> genomes	214.2	171.4	
1x10 <sup>10</sup> genomes	117.8	109.8	
5x10 <sup>10</sup> genomes	64.5	67.8	
2.5x10 <sup>10</sup> genomes	30.9	58.4	

rAAV-1 CMV-lacZ viruses were also injected into the muscle of C57BL6 mice and similar results were obtained. Collectively, these results suggested that AAV-1 based vector would be appropriate for both liver and muscle gene delivery.

### Example 5 - Neutralizing Antibodies Against AAV-1

Simple and quantitative assays for neutralizing antibodies (NAB) to AAV-1 and AAV-2 were developed with recombinant vectors. A total of 33 rhesus monkeys and 77 normal human subjects were screened.

#### A. Nonhuman Primates

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Wild-caught juvenile rhesus monkeys were purchased from Covance (Alice, Tex.) and LABS of Virginia (Yemassee, SC) and kept in full quarantine. The monkeys weighed approximately 3 to 4 kg. The nonhuman primates used in the Institute for Human Gene Therapy research program are purposefully bred in the United States from specific-pathogen-free closed colonies. All vendors are US Department of Agriculture class A dealers. The rhesus macaques are therefore not infected with important simian pathogens, including the tuberculosis agent, major simian lentiviruses (simian immunodeficiency virus and simian retroviruses), and cercopithecine herpesvirus. The animals are also free of internal and external parasites. The excellent health status of these premium animals minimized the potential for extraneous variables. For this study, serum was obtained from monkeys prior to initiation of any protocol.

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NAB titers were analyzed by assessing the ability of serum antibody to inhibit the transduction of reporter virus expressing green fluorescent protein (GFP) (AAV1-GFP or AAV2-GFP) into 84-31 cells. Various dilutions of antibodies preincubated with reporter virus for 1 hour at 37°C were added to 90% confluent cell cultures. Cells were incubated for 48 hours and the expression of green fluorescent protein was measured by FluoroImaging (Molecular Dynamics). NAB titers were calculated as the highest dilution at which 50% of the cells stained green.

Analysis of NAB in rhesus monkeys showed that 61% of animals tested positive for AAV-1; a minority (24%) has NAB to AAV-2. Over one-third of animals had antibodies to AAV-1 but not AAV-2 (i.e., were monospecific for AAV-1), whereas no animals were positive for AAV-2 without reacting to AAV-1. These data support the hypothesis that AAV-1 is endemic in rhesus monkeys. The presence of true AAV-2 infections in this group of nonhuman primates is less clear, since cross-neutralizing activity of an AAV-1 response to AAV-2 can not be ruled out. It is interesting that there is a linear relationship between AAV-2 NAB and AAV-1 NAB in animals that had both.

#### B. Humans

For these neutralization antibody assays, human serum samples were incubated at 56°C for 30 min to inactivate complement and then diluted in DMEM. The virus (rAAV or rAd with either lacZ or GFP) was then mixed with each serum dilution (20X, 400X, 2000X, 4000X, etc.) and incubated for 1 hour at 37°C before applied to 90% confluent cultures of 84-31 cells (for AAV) or Hela cells (for adenovirus) in 96-well plates. After 60 minutes of incubation at culture condition, 100 µl additional media containing 20% FCS was added to make final culture media containing 10% FCS.

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The result is summarized in Table 3.

Table 3

Adenovirus	AAV-1	AAV-2	# of samples	Percentage
-	-	-	41	53.2%
+	-	-	16	20.8%
•	+	-	0	0.0%
-	-	+	2	2.6%
-	+	+	2	2.6%
+	-	+	3	3.9%
. +	+	<u>-</u>	0 .	0.0%
+	+	+	13	16.9%
		Total	77	100%

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The human neutralizing antibodies against these three viruses seemed to be unrelated since the existence of neutralizing antibodies against AAV are not indications for antibodies against adenovirus. However, AAV requires adenovirus as helper virus, in most of the cases, the neutralizing antibodies against AAV correlated with the existence of neutralizing antibodies to adenovirus. Among the 77 human serum samples screened, 41% of the samples can neutralize the infectivity of recombinant adenovirus based on Ad5. 15/77 (19%) of serum samples can neutralize the transduction of rAAV-1 while 20/77 (20%) of the samples inhibit rAAV-2 transduction at 1 to 80 dilutions or higher. All serum samples positive in neutralizing antibodies for AAV-1 in are also positive for AAV-2. However, there are five (6%) rAAV-2 positive samples that failed to neutralize rAAV-1. In samples that are positive for neutralizing antibodies, the titer of antibodies also varied in the positive ones. The results from screening human sera for antibodies against AAVs supported the conclusion that AAV-1 presents the same epitome as that of AAV-2 to interact

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with cellular receptors since AAV-1 neutralizing human serums can also decrease the infectivity of AAV-2. However, the profile of neutralizing antibodies for these AAVs is not identical, there are additional specific receptors for each AAV serotype.

### Example 6 - Recombinant AAV Viruses Exhibit Tissue Tropism

The recombinant AAV-1 vectors of the invention and the recombinant AAV-2 vectors [containing the gene encoding human α1-antitrypsin (α1AT) or murine erythropoietin (Epo) from a cytomegalovirus-enhanced β-actin promoter (CB)] were evaluated in a direct comparison to equivalent copies of AAV-2 vectors containing the same vector genes.

Recombinant viruses with AAV-1 capsids were constructed using the techniques in Example 1. To make rAAV with AAV-1 virions, pAV1H or p5E18 (2/1) was used as the *trans* plasmid to provide Rep and Cap functions. For the generation of the rAAV based on AAV-2, p5E18(2/2) was used as the *trans* plasmid, since it greatly improved the rAAV yield. [Early experiments indicated similar *in vivo* performances of AAV-1 vectors produced with pAV1H and p5E19 (2/1). All subsequent studies used AAV-1 vectors derived from p5E18(2/1) because of the increased yield.]

Equivalent stocks of the AAV-1 and AAV-2 vectors were injected intramuscularly (5 x 10<sup>10</sup> genomes) or liver via the portal circulation (1 x 10<sup>11</sup> genomes) into immunodeficient mice, and the animals (four groups) were analyzed on day 30 for expression of transgene. See, Figs. 4A and 4B.

AAV-2 vectors consistently produced 10- to 50-fold more serum erythropoietin or α1-antitrypsin when injected into liver compared to muscle. (However, the AAV-1-delivered genes did achieve acceptable expression levels in the liver.) This result was very different from that for AAV-1 vectors, with which muscle expression was equivalent to or greater than liver expression. In fact, AAV-1 outperformed AAV-2 in muscle when equivalent titers based on genomes were administered.

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### Example 7 - Gene Delivery via rAAV-1

C57BL/6 mice (6- to 8-week old males, Jackson Laboratories) were analyzed for AAV mediated gene transfer to liver following intrasplenic injection of vector (i.e., targeted to liver). A total of 10<sup>11</sup> genome equivalents of rAAV-1 or rAAV-2 vector were injected into the circulation in 100 μl buffered saline. The first vector contained either an AAV-1 capsid or an AAV-2 capsid and expressed α1AT under the control of the chicken β-actin (CB) promoter. Day 28 sera were analyzed for antibodies against AAV-1 or AAV-2 and serum α1AT levels were checked. Animals were then injected with an AAV-1 or AAV-2 construct expressing erythropoietin (Epo, also under the control of the CB promoter). One month later sera was analyzed for serum levels of Epo. The following groups were analyzed (Figs. 5A-5D).

In Group 1, vector 1 was AAV-2 expressing a1AT and vector 2 was AAV-2 expressing Epo. Animals generated antibodies against AAV-2 following the first vector administration which prevented the readministration of the AAV-2 based vector. There was no evidence for cross-neutralizing the antibody to AAV-1.

In Group 2, vector 1 was AAV-1 expressing a1AT while vector 2 was AAV-1 expressing Epo. The first vector administration did result in significant a1AT expression at one month associated with antibodies to neutralizing antibodies to AAV-1. The animals were not successfully readministered with the AAV-1 Epo expressing construct.

In Group 3, the effectiveness of an AAV-2 vector expressing Epo injected into a naive animal was measured. The animals were injected with PBS and injected with AAV-2 Epo vector at day 28 and analyzed for Epo expression one month later. The neutralizing antibodies were evaluated at day 28 so we did not expect to see anything since they received PBS with the first vector injection. This shows that in naive animals AAV-2 is very efficient at transferring the Epo gene as demonstrated by high level of serum Epo one month later.

Group 4 was an experiment similar to Group 3 in which the animals originally received PBS for vector 1 and then the AAV-1 expressing Epo construct 28 days later. At the time of vector injection, there obviously were no antibodies to either

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AAV-1 or AAV-2. The AAV-1 based vector was capable of generating significant expression of Epo when measured one month later.

Group 5 is a cross-over experiment where the initial vector is AAV-2 expressing α1AT followed by the AAV-1 construct expressing Epo. The animals, as expected, were efficiently infected with the AAV-2 vector expressing α1AT as shown by high levels of the protein in blood at 28 days. This was associated with significant neutralizing antibodies to AAV-2. Importantly, the animals were successfully administered AAV-1 following the AAV-2 vector as shown by the presence of Epo in serum 28 days following the second vector administration. At the time of this vector administration, there was high level AAV-2 neutralizing antibodies and very low cross-reaction to AAV-1. The level of Epo was slightly diminished possibly due to a small amount of cross-reactivity. Group 6 was the opposite cross-over experiment in which the initial vector was AAV-1 based, whereas the second experiment was AAV-2 based. The AAV-1 vector did lead to significant gene expression of α1AT, which also resulted in high level AAV-1 neutralizing antibody. The animals were very efficiently administered AAV-2 following the initial AAV-1 vector as evidenced by high level Epo.

A substantially identical experiment was performed in muscle in which  $5 \times 10^{10}$  genomes were injected into the tibialis anterior of C57BL/6 mice as a model for muscle directed gene therapy. The results are illustrated in Figs. 6A-6D and are essentially the same as for liver.

In summary, this experiment demonstrates the utility of using an AAV-1 vector in patients who have pre-existing antibodies to AAV-2 or who had initially received an AAV-2 vector and need readministration.

### 25 Example 8 - Construction of Recombinant Viruses Containing AAV-1 ITRs

This example illustrates the construction of recombinant AAV vectors which contain AAV-1 ITRs of the invention.

An AAV-1 cis plasmid is constructed as follows. A 160 bp Xho-NruI AAV-1 fragment containing the AAV-1 5' ITR is obtained from pAV1-BL pAV1-BL was

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generated as described in Example 1. The Xho-NruI fragment is then cloned into a second pAV1-BL plasmid at an XbaI site to provide the plasmid with two AAV-1 ITRs. The desired transgene is then cloned into the modified pAV-1BL at the NruI and BamHI site, which is located between the AAV-1 ITR sequences. The resulting AAV-1 cis plasmid contains AAV-1 ITRs flanking the transgene and lacks functional AAV-1 rep and cap.

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Recombinant AAV is produced by simultaneously transfecting three plasmids into 293 cells. These include the AAV-1 cis plasmid described above; a trans plasmid which provides AAV rep/cap functions and lacks AAV ITRs; and a plasmid providing adenovirus helper functions. The rep and/or cap functions may be provided in trans by AAV-1 or another AAV serotype, depending on the immunity profile of the intended recipient. Alternatively, the rep or cap functions may be provided in cis by AAV-1 or another serotype, again depending on the patient's immunity profile.

In a typical cotransfection, 50 µg of DNA (cis:trans:helper at ratios of 1:1:2, respectively) is transfected onto a 15 cm tissue culture dish. Cells are harvested 96 hours post transfection, sonicated and treated with 0.5% sodium deoxycholate (37° for 10 min). Cell lysates are then subjected to 2-3 rounds of ultracentrifugation in a cesium gradient. Peak fractions containing rAAV are collected, pooled and dialyzed against PBS. A typical yield is 1 x 10<sup>13</sup> genomes/10<sup>9</sup> cells.

Using this method, one recombinant virus construct is prepared which contains the AAV-1 ITRs flanking the transgene, with an AAV-1 capsid. Another recombinant virus construct is prepared with contains the AAV-1 ITRs flanking the transgene, with an AAV-2 capsid.

All publications cited in this specification are incorporated herein by reference.

While the invention has been described with reference to a particularly preferred embodiments, it will be appreciated that modifications can be made without departing from the spirit of the invention. Such modifications are intended to fall within the scope of the claims.

### What is claimed is:

- 1. An isolated AAV-1 nucleic acid molecule comprising a sequence selected from the group consisting of:
  - (a) SEQ ID NO: 1;
  - (b) a DNA sequence complementary to SEQ ID NO: 1;
  - (c) cDNA complementary to (a) or (b); and
  - (d) RNA complementary to any of (a) to (c).
- 2. A nucleic acid molecule comprising an AAV-1 inverted terminal repeat (ITR) sequence selected from the group consisting of:
  - (a) nt 1 to 143 of SEQ ID NO: 1;
  - (b) nt 4576 to 4718 of SEQ ID NO: 1;
  - (c) a nucleic acid sequence complementary to (a) or (b); and
  - (d) a functional fragment of (a), (b), or (c).
- 3. A recombinant vector comprising a 5' AAV-1 inverted terminal repeat (ITR) and a selected transgene, wherein said ITR has the sequence selected from the group consisting of:
  - (a) nt 1 to 143 of SEQ ID NO: 1;
  - (b) a nucleic acid sequence complementary to (a); and
  - (c) a functional fragment of (a) or (b).
- 4. The recombinant vector according to claim 3, wherein said vector further comprises a 3' AAV-1 ITR.

- 5. A recombinant vector comprising a 3' AAV-1 inverted terminal repeat (ITR) and a selected transgene, wherein said ITR has the sequence selected from the group consisting of:
  - (a) nt 4576 to 4718 of SEQ ID NO: 1;
  - (b) a nucleic acid sequence complementary to (a); and
  - (c) a functional fragment of (a) or (b).
- 6. The recombinant vector according to claim 5, wherein said vector further comprises a 5' AAV-1 ITR.
- 7. The recombinant vector according to any of claims 3-6, wherein said vector further comprises AAV-1 capsid proteins having the sequence of SEQ ID NO: 13, 15 or 17 or functional fragments thereof.
- 8. The recombinant vector according to any of claims 3-6, wherein said vector further comprises adenovirus sequences.
- 9. A recombinant vector comprising an AAV-1 P5 promoter having the sequence of nt 236 to 299 of SEQ ID NO: 1 or a functional fragment thereof.
- 10. A nucleic acid molecule encoding AAV-1 helper functions, said molecule comprising an AAV rep coding region and an AAV cap coding region, wherein said cap coding region comprises at least one member is selected from the group consisting of:
  - (a) vp1, nt 2223 to 4431 of SEQ ID NO: 1;
  - (b) vp2, nt 2634 to 4432 of SEQ ID NO: 1; and
  - (c) vp3, nt 2829 to 4432 of SEQ ID NO: 1.

- 11. A nucleic acid molecule encoding AAV-1 helper functions, said molecule comprising an AAV rep coding region and an AAV cap coding region, wherein said rep coding region comprises an AAV-1 rep coding region comprising at least one member selected from the group consisting of:
  - (a) rep 78, nt 335 to 2304 of SEQ ID NO: 1;
- (b) rep 68, nt 335 to 2272 of SEQ ID NO: 1 or the cDNA corresponding thereto;
  - (c) rep 52, nt 1007 to 2304 of SEQ ID NO: 1; and
- (d) rep 40, nt 1007 to 2272 of SEQ ID NO: 1 or the cDNA corresponding thereto.
- 12. A host cell transduced with a recombinant viral vector according to any of claims 3-6.
- 13. A host cell transduced with a nucleic acid molecule according to any of claims 1, 2, 10 or 11.
- 14. A host cell stably transduced with an AAV-1 P5 promoter having the sequence of nt 236 to 299 of SEQ ID NO: 1.
- 15. A pharmaceutical composition comprising a carrier and a virus comprising the vector according to any of claims 3-6.
- 16. A pharmaceutical composition comprising a carrier and a virus comprising the vector according to claim 7.
- 17. A pharmaceutical composition comprising a carrier and a virus comprising the vector according to claim 8.

- 18. A method for AAV-mediated delivery of a transgene comprising the step of delivering to a host cell an AAV virion which comprises:
- (a) a capsid comprising at least one capsid protein encoded by an AAV-1 cap gene; and
- (b) a DNA molecule comprising a transgene under the control of regulatory sequences directing its expression.
- 19. A method for AAV-mediated delivery of a transgene to a host comprising the steps of:
- (a) assaying a sample from the host to determine the presence of neutralizing antibodies specific against any serotype of AAV; and
  - (b) delivering to the host an AAV virion which comprises:
- (i) a capsid comprising at least one capsid protein encoded by a cap gene of an AAV serotype against which the host has no antibodies as determined in step (a); and
- (ii) a DNA molecule comprising a transgene under the control of regulatory sequences directing its expression.
- The method according to claim 19, comprising the additional step of repeating steps (a) and (b).
- 21. Use of an AAV virion which comprises a capsid comprising (a) at least one capsid protein encoded by a cap gene of an AAV serotype against which the host has antibodies, and (b) a DNA molecule comprising a transgene operably linked to regulatory sequences directing its expression,

in the preparation of a medicament for delivery of a transgene to a host, wherein said host has no preexisting neutralizing antibodies against the AAV serotype of said cap gene.

- 22. A method for delivery of a transgene comprising the step of delivering to a host cell a recombinant virus comprising a recombinant vector according to any of claims 3-8.
- 23. A method for producing a selected gene product comprising the steps of transfecting a mammalian cell with the molecule according to claim 1 or a functional fragment thereof and culturing said cell under conditions suitable to express said gene product.

### FIG 1A

AAV-2	ttgcccactccctctctgcgcgctcgctcgctcggtggggcctgcggaccaaaggtccgcgg.gcgc	60 60 60
2-1144	Rep binding site  agacggcagagctctgctctgccggccccaccgagcgagc	120 120 120
7	?RS™	
AAV-1	ggcaactccatcactaggggtaaTCGCGAAGCGCCTCCCACGCTGCCGCGTCAGCGCTGA .cTG.AGcct.GGTG.AG	180 163 163
	E box/USF	
2-1744	CGTAAATTACGTCATAGGGGAGTGGTCCTGTATTAGCTGTCACGTGAGTGCTTTTGCGAGAGAG	237 222 222
	YY1 P <u>5/TATA</u>	
77177	GACATTTTGCGACACCACGTGGCCATTTAGGGTATATATGGCCGAGTGAGCGAGC	297 282 282
	70.460	
	YY1/p5 RNA Rep_78/68 CTCCATTTTGAC-CGCGAAATTTGAACGAGCAGCCATGCCGGGCTTCTACGAGATCG	356
7 7 77 2	AG G GGCCGT	342
AAV-6		341
3377 O	TGATCAAGGTGCCGAGCGACCTGGACGAGCACCTGCCGGGCATTTCTGACTCGTTTGTGATC.CTGTCAGCAGC	416 402 401
		476
AAV-1	GCTGGGTGGCCGAGAAGGAATGGGAGCTGCCCCCGGATTCTGACATGGATCTGAATCTGA	4 6 2
AAV-2	ATG.ATG.A	461
		536
AAV-1	TTGAGCAGGCACCCTGACCGTGGCCGAGAAGCTGCAGCGCGACTTCCTGGTCCAATGGC	522
AAV-2	5	521
		596
AAV-	GCCGCGTGAGTAAGGCCCCGGAGGCCCTCTTCTTTGTTCAGTTCGAGAAGGGCGAGTCCT	582
AAV-2	5	581
		65
AAV-	ACTTCCACCTCCATATTCTGGTGGAGACCACGGGGGTCAAATCCATGGTGCTGGGCCGCTA.GCG.GCACGTTAT.	642
AAV-	6	64
222	TO THE PETER CALLED A CONTROL OF THE	71
2.2.17	C C A. A. A. GA. T GA T	, 0.
AAV-	6	70:
7 7 T	1 TGCCCAACTGGTTCGCGGTGACCAAGACGCGTAATGGCGCCGGAGGGGGGAACAAGGTGG	77
AAV-	2 A	70.
1 12 1 V		76

# FIG 1B

7 7 T T T T	Ψ	836 822 821
2217 0	P19/TATA P19 RNA CGTGGACTAACATGGAGGAGTATATAAGCGCCTGTTTGAACCTGGCCGAGCGCAAACGGC	896 882 881
	TCGTGGCGCAGCACCTGACCCACGTCAGCCAGACCCAGGAGCAGAACAAGGAGAATCTGA .GTGGTCGGAA	956 942 941
	ACCCCAATTCTGACGCGCCTGTCATCCGGTCAAAAACCTCCGGGTCAAAAACCTCCGGTCAAAAACCTCCGGTCAAAAAACCTCCGGTCAAAAAACCTCCGGTCAAAAAACCTCCGGTCAAAAAACCTCCGGTCAAAAAACCTCCGGTCAAAAAACCTCCGGTCAAAAAACCTCCGGTCAAAAAACCTCCGGTCAAAAAACCTCCGGTCAAAAAACCTCCGGTCAAAAAACCTCCGGTCAAAAAACCTCCGGTCAAAAAACCTCCGGTCAAAAAACCTCCGGTCAAAAAACCTCCGGTCAAAAAACCTCCGGTCAAAAAAACCTCCGGTCAAAAAACCTCCGGTCAAAAAACCTCCGGTCAAAAAACCTCCGGTCAAAAAAAA	1016 1002 1001
AAV-1	TCGGGTGGCTGGTGGACCGGGGCATCACCTCCGAGAAGCAGTGGATCCAGGAGGACCAGG	1076 1062 1061
3317 0	CCTCGTACATCTCCTTCAACGCCGCTTCCAACTCGCGGTCCCAGATCAAGGCCGCTCTGGATGCATCT	1136 1122 1121
****	ACAATGCCGGCAAGATCATGGCGCTGACCAAATCCGCGCCCGACTACCTGGTAGGCCCCG	1196 1182 1181
	CTCCGCCCGCGGACATTAAAACCAACCGCATCTACCGCATCCTGGAGCTGAACGGCTACG AGCGTG.ATCC.GTGTTAAATTAAG	1256 1242 1241
	AACCTGCCTACGCCGGCTCCGTCTTTCTCGGCTGGGCCCAGAAAAGGTTCGGGAAGCGCA  T.CCAA.T.G.CT	1316 1302 1301
	ACACCATCTGGCTGTTTGGGCCGGCCACCACGGGCAAGACCAACATCGCGGAAGCCATCG	
-VAA -VAA	1 CCCACGCCGTGCCCTTCTACGGCTGCGTCAACTGGACCAATGAGAACTTTCCCTTCAATG 2A.T	1421
AAV- AAV-	1 ATTGCGTCGACAAGATGGTGATCTGGTGGGAGGGGGGAAGATGACGGCCAAGGTCGTGG 2 .CT	1481
AAV-	1 AGTCCGCCAAGGCCATTCTCGGCGGCAGCAAGGTGCGCGTGGACCAAAAGTGCAAGTCGT 2GAAA	154
AAV-	1 CCGCCCAGATCGACCCCACCCCGTGATCGTCACCTCCAACACATGTGCGCCGTGA	

### FIG 1C

	TTGACGGGAACAGCACCTTCGAGCACCAGCAGCAGCGGTGGAGCAGCAGCAGCAGCAGCAGCAGCAGCAGCAGCAGCAG	1676 1662 1661
nn** 1	THE A A CHECK COCCCCTCTCCACCATGACTTTGGCAAGGTGACAAAGCAGGAAGTCAAAG	1740
3 3 1 7 C	TTGAACTCACCCGCCGTCTGGACCATGACTCTCACC	1722 1721
	AGTTCTTCCGCTGGGCGCAGGATCACGTGACCGAGGTGGCGCATGAGTTCTACGTCAGAA .CTGAAGTTAAA.	1796 1782 1781
AAV-6		
	P40/TATA  AGGGTGGAGCCAACAAAAGACCCGCCCCCGATGACGCGGATAAAAGCGAGCCCAAGCGGG	1012
	P40 RNA	1916
0	P40 RNA CCTGCCCTCAGTCGCGGATCCATCGACGTCAGACGCGGAAGGAGCTCCGGTGGACTTTG	1000
•	· · · · · · · · · · · · · · · · · · ·	
AAV-1	TO SEE A COUNCIL A A A A A A TOTTOTOTO A COGGGGCATGCTT CAGATGCTGTTT CCCT	1976 1959
	T AA I	100
AAV-6		
	GCAAGACATGCGAGAGAATGAATCAGAATTTCAACATTTGCTTCACGCACG	2025
AAV-6	A	2021
AAV-1	ACTGTTCAGAGTGCTTCCCCGGCGTGTCAGAATCTCAACCGGTCGTCAGAAAGAGAGA	2093
AAV-6	5AT	20.0
AAV-1	CGTATCGGAAACTCTGTGCCATTCATCATCTGCTGGGGCGGGC	2153
	CGTATCGGAAACTCTGTGCCATTCATCATCTGCTGGGGGGGG	
	Rep 78 stop	
	1 CGGCCTGCGATCTGGTCAACGTGGACCTGGATGACTGTGTTTCTGAGCAATAAATGACTT	
AAV-	2 .T	
	$\nabla$ VP1 $\nabla$ Rep 68:	stop
AAV-	V VP1 1 AAACCAGGTATGGCTGCCGATGGTTATCTTCCAGATTGGCTCGAGGACAACCTCTCTGAG	3 2273 A 2253
-VAA -VAA	AAACCAGGTATGGCTGCCGATGGTTATCTTGCAGATTGGTGGTATCTTGCAGATTGGTGGTATCTTGCAGATTGGTGGTTATCTTGCAGATTGGTGGTTATCTTGCAGATTGGTGGTTATCTTGCAGATTGGTGGTTATCTTGCAGATTGGTGGTTATCTTGCAGATTGGTGGTTATCTTGCAGATTGGTGGTTATCTTGCAGATTGGTGGTTATCTTGCAGATTGGTGGTTATCTTGCAGATTGGTGGTTATCTTGCAGATTGGTGGTTATCTTGCAGATTGGTGGTTATCTTGCAGATTGGTGGTTATCTTGCAGATTGGTGGTTATCTTGCAGATTGGTGGTTATCTTGCAGATTGGTGGTTATCTTGCAGATTGGTGGTTATCTTGCAGATTGGTGGTTATCTTGCAGATTGGTGTGTATCTTGCAGATTGGTGTGTATCTTGCAGATTGGTGTGTATCTTGCAGATTGGTGTGTATCTTGCAGATTGGTGTGTATCTTGCAGATTGGTGTGTATCTTGCAGATTGGTGTGTATCTTGCAGATTGGTGTGTATCTTGCAGATTGGTGTATCTTGCAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGATGGTGTATCTTGGAGATTGGTGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGTATCTTGGAGATTGGTGATTGGAGATTGGTGATGGTGATGGTGATGGTATTGGAGATTGGTGTATGGTATTGGAG	G 2258
	1 GGCATTCGCGAGTGGTGGGACTTGAAACCTGGAGCCCGAAGCCCAAAGCCAACCAGCA 2AAA.ACA.GC.CCC.AACCAAGCGC	
AAV-	6	. 2010
71 TO T	1 AAGCAGGACGACGGCCGGGGTCTGGTGCTTCCTGGCTACAAGTACCTCGGACCCTTCAA	C 2393
-VAA		
2 32 1 V	2 C.TA	. 2318

#### FIG 1D

DDV-2	GGACTCGACAAGGGGGGGGCCCGTCAACGCGGCGGCGGCCCCTCGAGCACGACAAGAGAACA	2433
	CCCTACCACCTCAAAGCGGGTGACAATCCGTACCTGCGGTATAACCACGCCGAC	2513
2-1744	GG.CAGCACCAAC	2493
AAV-6	A.AGCGTGCGT	2498
ת א זו <i>ת</i> ת	GCCGAGTTTCAGGAGCGTCTGCAAGAAGATACGTCTTTTGGGGGCAACCTCGGGCGAGCA	2573
	GA	2553
AAV-2	C	2558
AAV-6		2633
AAV-1	GTCTTCCAGGCCAAGAAGCGGGTTCTCGAACCTCTCGGTCTGGTTGAGGAAGGCGCTAAG	2613
AAV-2		2618
AAV-6	T.TT	2010
	VP2	2602
AAV-1	ACGCTCCTGGAAAGAACGTCCGGTAGAGCAGTCGCCACAAGAGCCAGACTCCTCCTCG	2673
AAV-2	GAGA.GCTTGTG	
AAV-6	TGAC.TGGACAA	2070
. מממ	GGCATCGGCAAGACAGCCCAGCAGCCCGCTAAAAAAGAGACTCAATTTTGGTCAGACTGGC	2753
7777-2	A C A G G	2/33
AAV-6	T	2738
		2813
AAV-1	GACTCAGAGTCAGTCCCCGATCCACAACCTCTCGGAGAACCTCCAGCAACCTCTCGGAGAACCTCTCAGCAACCTCAACAA	2793
AAV-2	TGCCCAA	2798
AAV-6		
	VP3	
AAV-1	GTGGGACCTACTACAATGGCTTCAGGCGGTGGCGCACCAATGGCAGACAATAACGAAGGC	2873
AAV-2	C A A	2853
AAV-6		2858
		2022
AAV-1	GCCGACGGAGTGGGTAATGCCTCAGGAAATTGGCATTGCGATTCCACATGGCTGGGCGAC	2933 2913
AAV-2	AA	2913
7 7 7 7 A	AGAGTCATCACCACCAGCACCGCACCTGGGCCTTGCCCACCTACAATAACCACCTCTAC	2993
71 71 TT 1	A	2313
AAV-	5TCAA	2978
AAV-	L AAGCAAATCTCCAGTGCTTCAACGGGGGCCAGCAACGACAACCACTACTTCGGCTACAGC	3053
7.7.77	T $T$ $T$ $T$ $T$ $T$ $T$ $T$ $T$ $T$	3030
AAV-	5	3038
AAV-	L ACCCCTGGGGGTATTTTGATTTCAACAGATTCCACTGCCACTTTTCACCACGTGACTGG	3000
AAV-	2T	2020
	5TC	
n n t i	CAGCGACTCATCAACAACAATTGGGGATTCCGGCCCAAGAGACTCAACTTCAAACTCTTC	3173
T T T T T - 1	2 NN	2120
MAY T	6	3158
AAV-	1 AACATCCAAGTCAAGGAGGTCACGACGAATGATGGCGTCACAACCATCGCTAATAACCTT	323
AAV-	2 m a CAC.TACGGGTCT.	3210
		3218

# FIG 1E

	ACCAGCACGGTTCAAGTCTTCTCGGACTCGGAGTACCAGCTTCCGTACGTCCTCGGCTCT	12/0
AAV-1	GCGCACCAGGGCTGCCTCCCTCCGTTCCCGGCGGACGTGTTCATGATTCCGCAATACGGC	3353
AAV-2 AAV-6	A	3398
3 5 5 7 O	C III	3473 3450 3458
7 7 7 7 A	GAAGTGCCTTTCCACAGCAGCTACGCGCACAGCCAGACGTGCAGCAGCAGCGAGCG	3533 3510 3498
2217	C	3593 3570 3578
3 3 5 7 7	AACAAGGACTTGCTGTTTAGCCGTGGGTCTCCAGCTGGCATGTCTGTTCAGCCCAAAAAC C.GTCAAGGC.T.ATCT.AG.CCGGAG.GAGATCGG.ACT.T.GG	3653 3630 3638
7777.7	TGGCTACCTGGACCCTGTTATCGGCAGCAGCGCGTTTCTAAAACAAAACAGACAACAACTCCAA.CAGTCTG.GT	3713 3690 3698
AAV-1	A DODGO A THEFT A COMOCA CHOCATOCTTCA A A ATATA ACCTCA ATGGGCGTGA ATCCATC	3,30
2 2 2 7 7	ATCAACCCTGGCACTGCTATGGCCTCACACAAAGACGACGAAGACAAGTTCTTTCCCATG C.G.G.TGGC.CAAGCGTATTCA.	3833 3810 3818
AAV-	1 AGCGGTGTCATGATTTTTGGAAAAGAGAGCGCCGGAGCTTCAAACACTGCATTGGACAAT 2GTC.CCGGC.AGT.A.AGAAAATGTGAACA.TAG 6	3878
2 2 7 7	1 GTCATGATTACAGACGAAGAGGAAATTAAAGCCACTAACCCTGTGGCCACCGAAAGATTT 2	3953 3930 3938
AAV- AAV-	1 GGGACCGTGGCAGTCAATTTCCAGAGCAGCAGCACAGACCCTGCGACCGGAGATGTGCAT 2TT.TAT.TACCCAGAG.C.AG.ATCC	3998
AAV-	1 GCTATGGGAGCATTACCTGGCATGGTGTGGCAAGATAGAGACGTGTACCTGCAGGGTCCC 2 A.ACAAC.TTC.TACGCTT	

### FIG 1F

	ATTTGGGCCAAAATTCCTCACACAGATGGACACTTTCACCCGTCTCCTCTTATGGGCGGCCAGAGCTACC	4110
AAV-1	TTTGGACTCAAGAACCCGCCTCCTCAGATCCTCATCAAAAACACGCCTGTTCCTGCGAATCTACTATGCGA	
7 7 T T 2	CCTCCGGCGGAGTTTTCAGCTACAAAGTTTGCTTCATTCA	4253 4230 4238
7 77 77 77	CA-AGTGAGTGTGGAAATTGAATGGGAGCTGCAGAAAGAAACAGCAAGCGCTGGAATCCCGCGCG	4312 4290 4297
3 3 TT 0	CGAAGTGCAGTACACATCCAATTATGCAAAATCTGCCAA-CGTTGATTTTACTGTGGACAA.TTCCAACGTTTGCTTTC	4371 4350 4356
3 3 TT 0	ACAATGGACTTTATACTGAGCCTCGCCCCATTGGCACCCGTTACCTTACCCGTCCCCTGTCTCG.GT.AA.AGTAAT	4431 4410 4416
	VP1-3 stopPolyA signalAATTACGTGTTAATCAATAAACCGGTTGATTCGTTTCAGTTGAACTTTGGTCTCCTGTCCG.T	. 7110
	TTCTTATCTTATC-GGTTACCATGGTTAT-AGCTTACACATTAACTGCTTGGTTGCGC .TC.TTATCCGTAGAAGT.GC.TGG.G.GAA.CATTAATCA.CA.C-C.GAA	4220
	TTCGCGATAAAAGACTTACGTCATCGGGttacccctagtgatggagttgcccactccctc ACTA.A.gg.a	4370
カカオエーク	tctgcgcgctcgctcggtggggccggcagagcagagctctgccgtctgcggacctt .cacacagccaggca.gc.c.gg	
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---aav-1 ~

AAV-1 TR

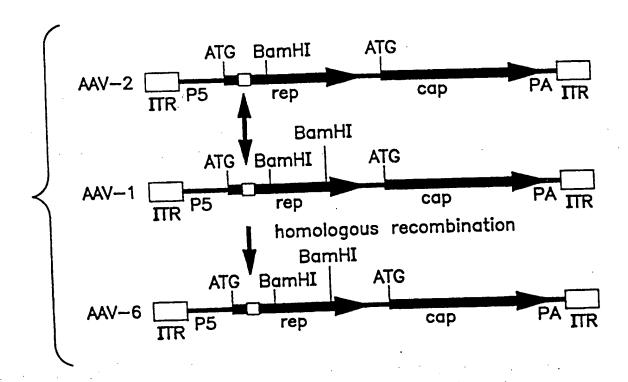


FIG. 3A

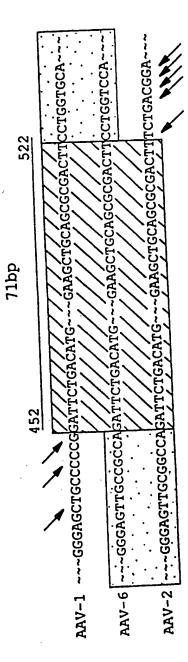


FIG. 3B

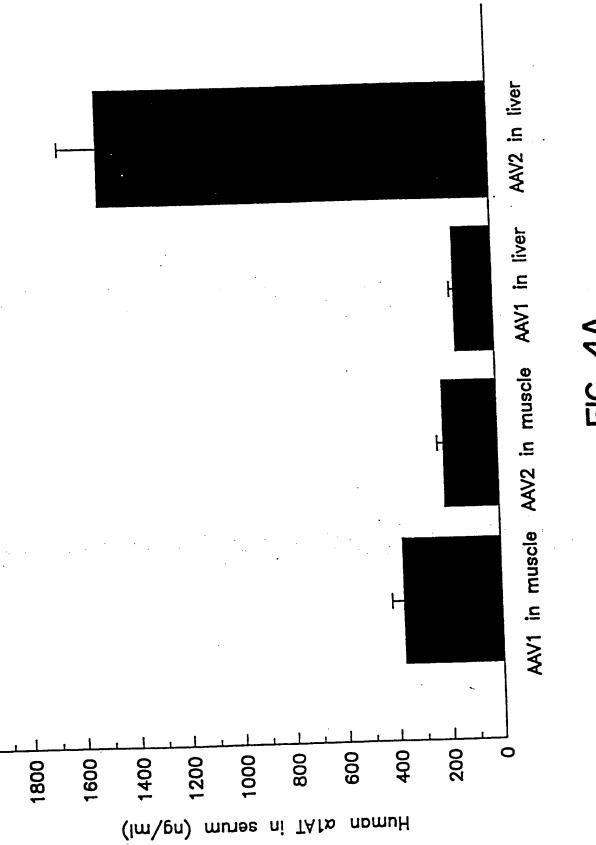
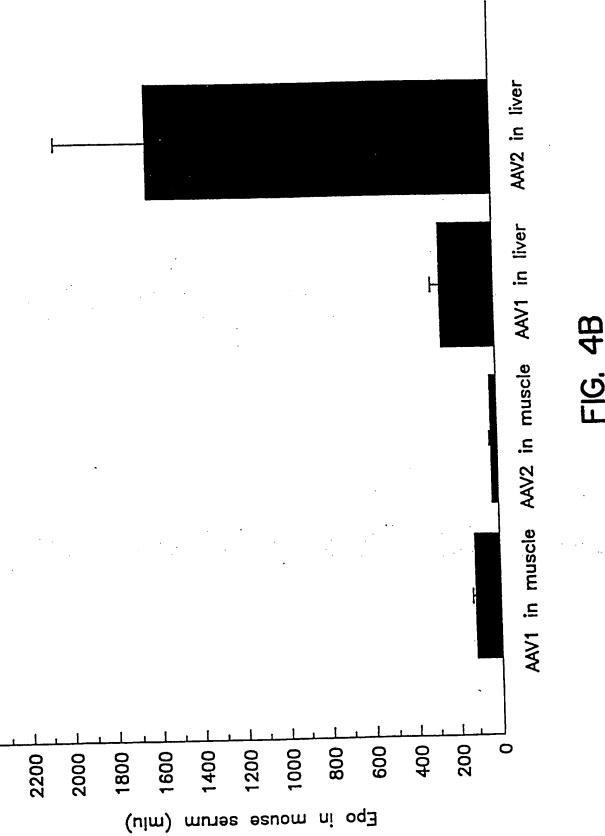
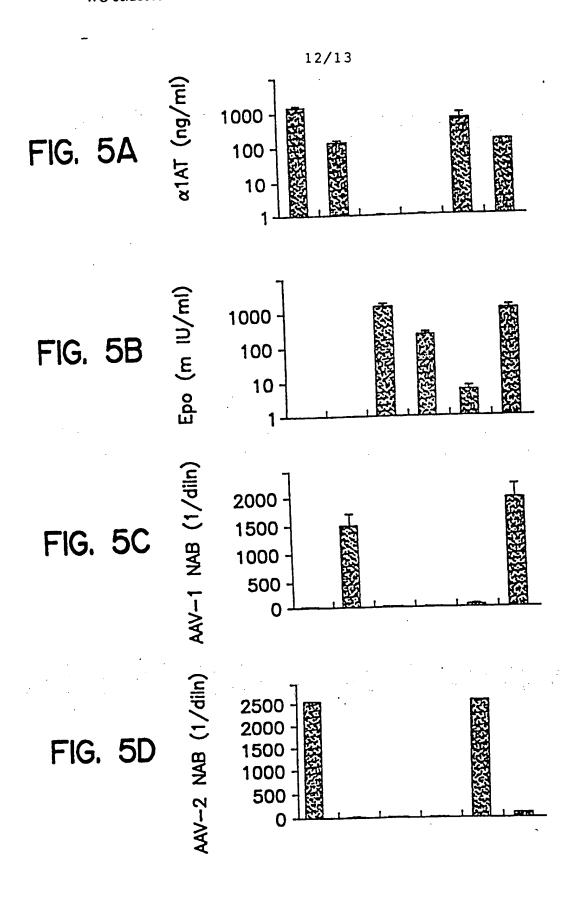
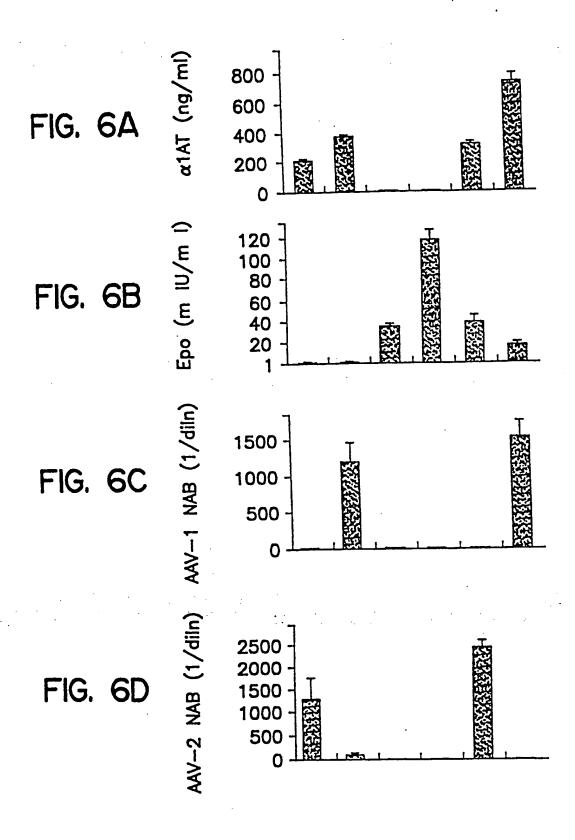


FIG. 4A





Group	1	2	3	4	5	6
Vector1- alAT	AAV2	AAV1	PBS	PBS	AAV2	AAV1
Vector2-EPO	AAV2	AAV1	AAV2	AAV1	AAV1	AAV2



1	Group	1	2	3	4	5_	6
	Vector1 - α1AT	AAV2	AAV1	PBS	PBS	AAV2	AAV1
	Vector2-EPO	AAV2	AAV1	AAV2	AAV1	AAV1	AAV2

PCT/US99/25694 WO 00/28061

SEQUENCE LISTING

<110> Wilson, James M. Xiao, Weidong The Trustees of the University of Pennsylvania

<120> Adeno-Associated Virus Serotype I Nucleic Acid Sequences, Vectors and Host Cells Containing Same

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<151> 1998-11-05

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1

1

-																
gtg	atc	aag	gtg	ccg	agc	gac	ctg	gac	gag	cac	ctg	ccg	ggc	att	tct	403
Val	Ile	Lys	Val	Pro	Ser	Asp	Leu	Asp	Glu	His	Leu	Pro	Gly	Ile	Ser	
		10					15					20				
			· · · .													
gac	tcg	ttt	gtg	agc	tgg	gtg	gcc.	gag	aag	gaa	tgg	gag	ctg	ccc	ccg	451
Asp	Ser	Phe	Val	Ser	Trp	Val	Ala	Glu	Lys	Glu	Trp	Glu	Leu	Pro	Pro	
	25					30					35					
gat	tct	gac	atg	gat	ctg	aat	ctg	att	gag	cag	gca	ccc	ctg	acc	gtg	499
Asp	Ser	Asp	Met	Asp	Leu	Asn	Leu	Ile	Glu	Gln	Ala	Pro	Leu	Thr		
40					45					50					55	
gcc	gag	aag	ctg	cag	cgc	gac	ttc	ctg	gtc	caa	tgg	cgc	cgc	gtg	agt	547
Ala	Glu	Lys	Leu	Gln	Arg	Asp	Phe	Leu	Val	Gln	Trp	Arg	Arg	Val	Ser	
				60		•			65					70		
aag	gcc	ccg	gag	gcc	ctc	ttc	ttt	gtt	cag	ttc	gag	aag	ggc	gag	tcc	595
Lys	Ala	Pro	Glu	Ala	Leu	Phe	Phe	Val	Gln	Phe	Glu	Lys	Gly	Glu	Ser	
-			75					80					85			
tac	ttc	cac	ctc	cat	att	ctq	qtq	gag	acc	acg	ggg	gtc	aaa	tcc	atg	643
Tur	Phe	His	Leu	His	Ile	Leu	Val	Glu	Thr	Thr	Gly	Val	Lys	Ser	Met	
ryr	·	90					9.5				•	100	-			٠.
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~+~	ata		cac	ttc	cta	agt	cad	att	agg	σac	aaq	cta	ata	cag	acc	691
grg	tan	ggc	n ~~	Dha	Lau	Ser	Gln	Tle	Ara	Asn	LVS	Leu	Val	Gln	Thr	
vaı			AIG	PHE	ьеи	110			9	7100	115					
	105	)				110										
				250	~ ~ ~	660	300	cta		aac	· taa	ttc	aca	gtg	acc	739
atc	tac	: cgc	999	alc	gag	D=0	Thr	tou	Dro	Asn	Trn	Phe	Δla	Val	Thr	
		Arg	GIY	ire			1111	, hea	110	130		2	2		135	
120	ı				125	1				130	,				100	
											. ~+ ~	ata	~~~		tac	787
aag	acq	g cgt	aat	ggc	gcc	gga	999	999	aac . nam	. aag	, 4.1	y y cy	ner	gag	Cvs	, , ,
Lys	Thi	r Arç	Asn			GIZ	GIA	, G13			o vai	. vai	Ash	Glu 150		
		• • •		140					145	<u>,</u>	ă.	-		150		
													o to		taa	835
tac	ato	c cc	aac	: tac	cto	cto	- L c/c c	aaç	act	. caç		gay		, cla	tgg	030
Ty	: Il	e Pro			Let	ı Lev	Pro			GII	1 PIC	) GIU	165	: :	Trp	
			155					160	,				10	,		
								نطيا			- +-·	. <b></b>				883
gc	g tg	g act	t aad	ato	ggaq	g gaq	g tat	ata 	ago	gc	cgt - c		, aac	. Tan	gcc	505
Ala	a Tr			n Met	: Gl	ı Glı			e Sei	c Ala	a Cys			т тек	Ala	
		17	0				175	<b>5</b>				180	,	,		
																931
ga	g cg	c aa	a cg	g cto	gt	g gc	g cad	g ca	cct	g aco	c cad	gto	ago	caç	acc	331
Gl	ı Ar	g Ly	s Ar	g Lei	ı Val			n Hi	s Le	ı Th			Se	E GIT	Thr	
	18	5				19	j .				195	5				

cag ( Gln (				- •		_					-		Pro	•	979
atc (														-	1027
gtg (								_			_		-	-	1075
gcc 1						-	=			_			-		1123
aag ( Lys )					_		_		_		-				1171
gcg ( Ala 1 280	_		_	-			-	_			-				1219
aac ( Asn )			-		_		_				_		-		1267
gcc (		-					-	_					-	_	1315
aac Asn '															1363
gcg (											-	-			1411
Thr 3							-	-	_	-	_	_			1459
tgg Trp'													-	_	1507

_																
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His	Asp	Phe	Gly	Lys	Val	Thr	Lys	Gln	Glu	Val	Lys	Glu	Phe	Phe	Arg	
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Trp	Ala	Gln	Asp	His	Val	Thr	Glu	Val	Ala	His	Glu	Phe	Tyr	Val	Arg	
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Asp	Cys		Glu	Cys	Phe	Pro	Gly	Val	Ser	Glu	Ser	Gln	Pro	Val	Val	
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_																
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_	gat Asp	-	_	-				taa	atga	actta	aaa o	cago	Me		et gcc La Ala	2231
-	ggt Gly				-								-			2279
-	gag Glu 645			_	_				-	_	-			-		2327
_	caa Gln		_	-	-					-					-	2375
	ctc Leu							-	_				-			2423
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	gcg		•		-		_					-	-	-		2519
	cag Gln 725	Glu	_	-		_	-									2567
_	gca Ala	-		-	_	-	_		-		-					2615
-	gag Glu	-		_	-					-						2663

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Gln	Ser	Pro	Gln	Glu	Pro	Asp	Ser	Ser	Ser	Gly	Ile	Gly	Lys	Thr	Gly	
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	Gln															•
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-	Ser	-		-												
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act	gct	ata	gga	cct	act	aca	atα	act	tca	aac	aat	aac	αca	сса	ato	2855
-	Ala															
820		***	Cly	110	825			,		830	0-7	,			835	
020					025					050					000	
002	gac	aat	aac	gaa	aac	acc	aac	ana	ata	aat	aat	acc	tca	gga	aat	2903
	Asp															2300
AIG	Yab	M311	Vall	840	Gry	AIG	Yab	OLY	845	Ory	7.511	710	501	850	AJII	
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taa	cat	tac	aat	tcc	aca	taa	cta	aac	gac	aga	atc	atc	acc	acc	age	2951
	His	•	-				_	-								2702
тър	urs	Суз	855	Ser	1111	TIP	пса	860	ДЭР	ALG	V 44 1	110	865	****	501	
			033		•			000		•			003			•
3.00	cgc	366	taa	acc	tta	CCC	200	tac	aat	220	cac	ctc	tac	aar	caa	2999
	Arg															2333
1111	ALY	870	rrp	YIG	Leu	FIO	875	LyL	7311	7311		880	1 y L	БуЗ	0111	
		670					0/5					000				
	tcc	- a +	aat	<b>t.c.</b>	266	~~~	~~~	200	336	a 2 c	220	C 2 C	t a c	ttc	aac	3047
		-														3047
116	Ser	ser	Ala	Ser	1111	-	Ald	Ser	ASII	Asp	895	nis	ıyı	FIIC	GIY	
	885					890					093					
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	agc										_					3033
_	Ser	Thr	Pro	Trp	_	Tyr	Phe	Asp	Pne		Arg	Pne	HIS	Cys		
900	۱	,			905					910					915	
		••			•			•		•	·					
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Ph∈	Ser	Pro	Arg	Asp	Trp	Gln	Arg	Leu		Asn	Asn	Asn	Trp		Phe	
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Arg	J Pro	Lys	Arg	Leu	Asn	Phe	Lys		Phe	Asn	Ile	Gln		Lys	Glu	
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														`.		
gto	acg	acg	aat	gat	ggc	gtc	aca	acc	atc	gct	aat	aac	ctt	acc	agc	3239
Va]	Thr	Thr	Asn	Asp	Gly	Val	Thr	Thr	Ile	Ala	Asn	Asn	Leu	Thr	Ser	
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	_		•														
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	-	-		Val		_	-	_			_						
		965					970			•		975		•	. •		
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				His													
	980					985	•				990			-		995	
	atq	att	cca	caa	tac	aac	tac	cta	acq	ctc	aac	aat	aac	agc	caa	qcc	3383
	_		-	Gln				-									
					1000	-	•			1005			-		1010		
									-								
	ata	gga	cat	tca	tcc	ttt	tac	tac	cta	gaa	tat	ttc	cct	tct	cag	ato	3431
			-	Ser				-	_	-							
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			•						. 020								
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	-	-	-	Gly						-						_	
	reu	-	1030	Gry	MSII	ASII		1035	FIIE	261	ıyı		1040	Giu	Giu	V G I	
			1030				•	1033				•	1040				•
				200	200	+ > 0	~~~	~~~	200		200	cta	<b>a</b> 36	caa	cta	ata	3527
				agc	-					_	_	_			_		3327
•			HIS	ser	ser	-		HIS	ser	GIN			Asp	ALG	ьец	Met	
		1045				٠.	1050				•	1055					5
														•			2575
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	-			agt													3623
	Gln	Ser	Gly	Ser		GIn	Asn	гàг	_		Leu	Phe	Ser			Ser	
					1080					1085					1090		
																	0.671
		_		atg													3671
	Pro	Ala	Gly	Met	Ser	Val	Gln			Asn	Trp	Leu	Pro	Gly	Pro	Cys	
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			•	•							•				•		
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	Tyr	Arg	Gln	Gln	Arg	Val	Ser	Lys	Thr	Lys	Thr	Asp	Asn	Asn	Asn	Ser	
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															`		
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	Ser	Ile	Ile	Asn	Pro	Gly	Thr	Ala	Met	Ala	Ser	His	Lys	Asp	Asp	Glu	
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Pro Arg Pro Ile Gly Thr Arg Tyr Leu Thr Arg Pro Leu

1350

1360

4440

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Glu Gln Ala Pro Leu Thr Val Ala Glu Lys Leu Gln Arg Asp Phe Leu 50 55 60

Val Gln Trp Arg Arg Val Ser Lys Ala Pro Glu Ala Leu Phe Phe Val 65 70 75 80

Gln Phe Glu Lys Gly Glu Ser Tyr Phe His Leu His Ile Leu Val Glu 85 90 95

Thr Thr Gly Val Lys Ser Met Val Leu Gly Arg Phe Leu Ser Gln Ile 100 105 110

Arg Asp Lys Leu Val Gln Thr Ile Tyr Arg Gly Ile Glu Pro Thr Leu 115 120 125

Pro Asn Trp Phe Ala Val Thr Lys Thr Arg Asn Gly Ala Gly Gly 130 135 140

Asn Lys Val Val Asp Glu Cys Tyr Ile Pro Asn Tyr Leu Leu Pro Lys Thr Gln Pro Glu Leu Gln Trp Ala Trp Thr Asn Met Glu Glu Tyr Ile Ser Ala Cys Leu Asn Leu Ala Glu Arg Lys Arg Leu Val Ala Gln His Leu Thr His Val Ser Gln Thr Gln Glu Gln Asn Lys Glu Asn Leu Asn Pro Asn Ser Asp Ala Pro Val Ile Arg Ser Lys Thr Ser Ala Arg Tyr Met Glu Leu Val Gly Trp Leu Val Asp Arg Gly Ile Thr Ser Glu Lys Gln Trp Ile Gln Glu Asp Glr. Ala Ser Tyr Ile Ser Phe Asn Ala Ala Ser Asn Ser Arg Ser Gln Ile Lys Ala Ala Leu Asp Asn Ala Gly Lys Ile Met Ala Leu Thr Lys Ser Ala Pro Asp Tyr Leu Val Gly Pro Ala Pro Pro Ala Asp Ile Lys Thr Asn Arg Ile Tyr Arg Ile Leu Glu Leu Asn Gly Tyr Glu Pro Ala Tyr Ala Gly Ser Val Phe Leu Gly Trp Ala Gln Lys Arg Phe Gly Lys Arg Asn Thr Ile Trp Leu Phe Gly Pro Ala Thr Thr Gly Lys Thr Asn Ile Ala Glu Ala Ile Ala His Ala Val Pro Phe Tyr Gly Cys Val Asn Trp Thr Asn Glu Asn Phe Pro Phe Asn Asp Cys Val Asp Lys Met Val Ile Trp Trp Glu Glu Gly Lys Met Thr Ala Lys Val Val Glu Ser Ala Lys Ala Ile Leu Gly Gly Ser Lys Val Arg 

Val Asp Gln Lys Cys Lys Ser Ser Ala Gln Ile Asp Pro Thr Pro Val 405 410 415

Ile Val Thr Ser Asn Thr Asn Met Cys Ala Val Ile Asp Gly Asn Ser
420 425 430

Thr Thr Phe Glu His Gln Gln Pro Leu Gln Asp Arg Met Phe Lys Phe 435 440 445

Glu Leu Thr Arg Arg Leu Glu His Asp Phe Gly Lys Val Thr Lys Gln 450 455 460

Glu Val Lys Glu Phe Phe Arg Trp Ala Gln Asp His Val Thr Glu Val 465 470 475 480

Ala His Glu Phe Tyr Val Arg Lys Gly Gly Ala Asn Lys Arg Pro Ala-485 490 495

Pro Asp Asp Ala Asp Lys Ser Glu Pro Lys Arg Ala Cys Pro Ser Val
500 505 510

Ala Asp Pro Ser Thr Ser Asp Ala Glu Gly Ala Pro Val Asp Phe Ala 515 520 525

Asp Arg Tyr Gln Asn Lys Cys Ser Arg His Ala Gly Met Leu Gln Met 530 540

Leu Phe Pro Cys Lys Thr Cys Glu Arg Met Asn Gln Asn Phe Asn Ile 545 550 555 560

Cys Phe Thr His Gly Thr Arg Asp Cys Ser Glu Cys Phe Pro Gly Val 565 570 575

Ser Glu Ser Gln Pro Val Val Arg Lys Arg Thr Tyr Arg Lys Leu Cys 580 585 590

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Cys Asp Leu Val Asn Val Asp Leu Asp Asp Cys Val Ser Glu Gln 610 615 620

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- Lys Ala Asn Gln Gln Lys Gln Asp Asp Gly Arg Gly Leu Val Leu Pro
- Gly Tyr Lys Tyr Leu Gly Pro Phe Asn Gly Leu Asp Lys Gly Glu Pro 50 55 60
- Val Asn Ala Ala Asp Ala Ala Ala Leu Glu His Asp Lys Ala Tyr Asp 65 70 75 80
- Gln Gln Leu Lys Ala Gly Asp Asn Pro Tyr Leu Arg Tyr Asn His Ala 85 90 95
- Asp Ala Glu Phe Gln Glu Arg Leu Gln Glu Asp Thr Ser Phe Gly Gly 100 105 110
- Asn Leu Gly Arg Ala Val Phe Gln Ala Lys Lys Arg Val Leu Glu Pro 115 120 125
- Leu Gly Leu Val Glu Glu Gly Ala Lys Thr Ala Pro Gly Lys Lys Arg 130 135 140
- Pro Val Glu Gln Ser Pro Gln Glu Pro Asp Ser Ser Ser Gly Ile Gly 145 150 155 160
- Lys Thr Gly Gln Gln Pro Ala Lys Lys Arg Leu Asn Phe Gly Gln Thr 165 170 175
- Gly Asp Ser Glu Ser Val Pro Asp Pro Gln Pro Leu Gly Glu Pro Pro 180 185 190
- Ala Thr Pro Ala Ala Val Gly Pro Thr Thr Met Ala Ser Gly Gly Gly 195 200 205
- Ala Pro Met Ala Asp Asn Asn Glu Gly Ala Asp Gly Val Gly Asn Ala 210 215 220
- Ser Gly Asn Trp His Cys Asp Ser Thr Trp Leu Gly Asp Arg Vàl Ile 225 230 235 240
- Thr Thr Ser Thr Arg Thr Trp Ala Leu Pro Thr Tyr Asn Asn His Leu

-		•		245					250					255	
Tyr	Lys	Gln	Ile 260	Ser	Ser	Ala	Ser	Thr 265	Gly	Ala	Ser	Asn	Asp 270	Asņ	His
Tyr	Phe	Gly 275	Туr	Ser	Thr	Pro	Trp 280	Gly	Tyr	Phe	Asp	Phe 285	Asn	Arg	Phe
His	Cys 290	His	Phe	Ser	Pro	Arg 295	Asp	Trp	Gln	Arg	Leu 300	Ile	Asn	Asn	Asn
Trp 305	Gly	Phe	Arg	Pro	Lys 310	Arg	Leu	Asn	Phe	Lys 315	Leu	Phe	Asn	Ile	Gln 320
Val	Lys	Glu	Val	Thr 325	Thr	Asn	Asp	Gly	Val 330	Thr	Thr	Ile	Ala	Asn 335	Asn
Leu	Thr	Ser	Thr 340	Val	Gln	Val	Phe	Ser 345	Asp	Ser	Glu	Tyr	Gln 350	Leu	Pro
Tyŗ	Val	Leu 355	Gly	Ser	Ala	Hıs	Gln 360	Gly	Cys	Leu	Pro	Pro 365	Phe	Pro	Ala
Asp	Val 370	Phe	Met	Ile	Pro	Gln 375	Tyr	Gly	Tyr	Leu	Thr 380	Leu	Asn	Asn	Glý
Ser 385	Gln	Ala	Val	Gly	Arg 390	Ser	Ser ,	Phe	Tyr	Cys 395	Leu	Glu	Tyr	Phe	Pro 400
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Glu	Ģlu	Val	Pro 420	Phe	His	Ser	Ser	Tyr 425	Ala	His	Ser	Gln	Ser 430	Leu	Asp
Arg	Leu	Met 435	Asn	Pro	Leu	Ile	Asp 440	Gln	Tyr	Leu	Tyr	Tyr 445	Leu	Asn	Arg
Thr	Gln 450	Asn	Gln	Ser	Gly	Ser 455	Ala	Gln	Asn	Lys	Asp 460	Leu	Leu	Phe	Ser
Arg 465	Gly	Ser	Pro	Ala	Gly 470	Met	Ser	Val	Gln	Pro 475	Lys	Asn	Trp	Leu	Pro 480
Gly	Pro	Cys	Туr	Arg 485	Gln	Gln	Arg	Val	Ser 490	Lys	Thr	Lys	Thr	Asp 495	Asn
Asn	Asn	ser	Asn	Phe	Thr	Trp	Thr	Gly	Ala	Ser	Lys	туr	Asn	Leu	Asn

\_ 500 505 510

Gly Arg Glu Ser Ile Ile Asn Pro Gly Thr Ala Met Ala Ser His Lys 515 520 525

Asp Asp-Glu Asp Lys Phe Phe Pro Met Ser Gly Val Met Ile Phe Gly 530 540

Lys Glu Ser Ala Gly Ala Ser Asn Thr Ala Leu Asp Asn Val Met Ile 545 550 555 560

Thr Asp Glu Glu Ile Lys Ala Thr Asn Pro Val Ala Thr Glu Arg
565 570 575

Phe Gly Thr Val Ala Val Asn Phe Gln Ser Ser Ser Thr Asp Pro Ala 580 585 590

Thr Gly Asp Val His Ala Met Gly Ala Leu Pro Gly Met Val Trp Gln
595 600 605

Asp Arg Asp Val Tyr Leu Gln Gly Pro Ile Trp Ala Lys Ile Pro His 610 615 620

Thr Asp Gly His Phe His Pro Ser Pro Leu Met Gly Gly Phe Gly Leu 625 630 635 640

Lys Asn Pro Pro Pro Gln Ile Leu Ile Lys Asn Thr Pro Val Pro Ala 645 650 655

Asn Pro Pro Ala Glu Phe Ser Ala Thr Lys Phe Ala Ser Phe Ile Thr 660 665 670

Gln Tyr Ser Thr Gly Gln Val Ser Val Glu Ile Glu Trp Glu Leu Gln 675 680 685

Lys Glu Asn Ser Lys Arg Trp Asn Pro Glu Val Gln Tyr Thr Ser Asn 690 695 700

Tyr Ala Lys Ser Ala Asn Val Asp Phe Thr Val Asp Asn Asn Gly Leu 705 710 715 720

Tyr Thr Glu Pro Arg Pro Ile Gly Thr Arg Tyr Leu Thr Arg Pro Leu
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<211> 1872

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act cag ccc gag ctg cag tgg gcg tgg act aac atg gag gag tat ata

528

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			tgt Cys										_		-		576
			cac His 195														624
			tct Ser												_		672
1			ctg Leu													_	720
			atc Ile												-	-	768
			tcg Ser														816
			gcg Ala 275													-	864
			gcg Ala														912
P			tac Tyr														960
			agg Arg									_			_	-	1008
			ggc Gly											-			1056
t	tc	tac	ggc	tgc	gtc	aac	tgg	acc	aat	gag	aac	ttt	ccc	ttc	aat	gat	1104

P <u>h</u> e	Туr	Gly 355	Cys	Val	Asn	Trp	Thr 360	Asn	Glu	Asn	Phe	Pro 365	Phe	Asn	Asp	
					gtg Val							_	-	_	_	1152
					gcc Ala 390								_		_	1200
					aag Lys											1248
					acc Thr							-			-	1296
					cag Gln							_				1344
					ctg Leu					-				_	_	1392
					ttc Phe 470											1440
					gtc Val											1488
					aaa Lys										gtc Val	1536
					tca Ser										-	1584
					aaa Lys											1632
ctg	ttt	ccc	tgc	aag	aca	tgc	gag	aga	atg	aat	cag	aat	ttc	aac	att	1680

Leu Phe Pro Cys Lys Thr Cys Glu Arg Met Asn Gln Asn Phe Asn Ile 545 550 555 560 tgc ttc acg cac ggg acg aga gac tgt tca gag tgc ttc ccc ggc gtg 1728 Cys Phe Thr His Gly Thr Arg Asp Cys Ser Glu Cys Phe Pro Gly Val 570 565 tca gaa tct caa ccg gtc gtc aga aag agg acg tat cgg aaa ctc tgt Ser Glu Ser Gln Pro Val Val Arg Lys Arg Thr Tyr Arg Lys Leu Cys 580 585 gcc att cat ctg ctg ggg cgg gct ccc gag att gct tgc tcg gcc Ala Ile His His Leu Leu Gly Arg Ala Pro Glu Ile Ala Cys Ser Ala 595 600 605 tgc gat ctg gtc aac gtg gac ctg gat gac tgt gtt tct gag caa taa 1872 Cys Asp Leu Val Asn Val Asp Leu Asp Asp Cys Val Ser Glu Gln 610 615 620 <210> 5 <211> 623

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Lys Glu Trp Glu Leu Pro Pro Asp Ser Asp Met Asp Leu Asn Leu Ile 35 40

Glu Gln Ala Pro Leu Thr Val Ala Glu Lys Leu Gln Arg Asp Phe Leu 50 60 - 55

Val Gln Trp Arg Arg Val Ser Lys Ala Pro Glu Ala Leu Phe Phe Val 65 70 75

Gln Phe Glu Lys Gly Glu Ser Tyr Phe His Leu His Ile Leu Val Glu 85 90

Thr Thr Gly Val Lys Ser Met Val Leu Gly Arg Phe Leu Ser Gln Ile 100 105 110

Arg Asp Lys Leu Val Gln Thr Ile Tyr Arg Gly Ile Glu Pro Thr Leu

1ε

115 120 125

Pro Asn Trp Phe Ala Val Thr Lys Thr Arg Asn Gly Ala Gly Gly 130 135 140 .

Asn Lys Val Val Asp Glu Cys Tyr Ile Pro Asn Tyr Leu Leu Pro Lys 145 150 155 160

Thr Gln Pro Glu Leu Gln Trp Ala Trp Thr Asn Met Glu Glu Tyr Ile 165 170 175

Ser Ala Cys Leu Asn Leu Ala Glu Arg Lys Arg Leu Val Ala Gln His 180 185 190

Leu Thr His Val Ser Gln Thr Gln Glu Gln Asn Lys Glu Asn Leu Asn 195 200 205

Pro Asn Ser Asp Ala Pro Val Ile Arg Ser Lys Thr Ser Ala Arg Tyr 210 215 220

Met Glu Leu Val Gly Trp Leu Val Asp Arg Gly Ile Thr Ser Glu Lys 225 230 235 240

Gln Trp Ile Gln Glu Asp Gln Ala Ser Tyr Ile Ser Phe Asn Ala Ala 245 250 255

Ser Asn Ser Arg Ser Gln Ile Lys Ala Ala Leu Asp Asn Ala Gly Lys 260 265 270

Ile Met Ala Leu Thr Lys Ser Ala Pro Asp Tyr Leu Val Gly Pro Ala 275 280 285

Pro Pro Ala Asp Ile Lys Thr Asn Arg Ile Tyr Arg Ile Leu Glu Leu 290 295 300

Asn Gly Tyr Glu Pro Ala Tyr Ala Gly Ser Val Phe Leu Gly Trp Ala 305 310 315 320

Gln Lys Arg Phe Gly Lys Arg Asn Thr Ile Trp Leu Phe Gly Pro Ala 325 330 335

Thr Thr Gly Lys Thr Asn Ile Ala Glu Ala Ile Ala His Ala Val Pro 340 345 350

Phe Tyr Gly Cys Val Asn Trp Thr Asn Glu Asn Phe Pro Phe Asn Asp 355 360 365

Cys Val Asp Lys Met Val Ile Trp Trp Glu Glu Gly Lys Met Thr Ala

**- 370 375 380** 

Lys Val Val Glu Ser Ala Lys Ala Ile Leu Gly Gly Ser Lys Val Arg 385 390 395 400

Val Asp Gln Lys Cys Lys Ser Ser Ala Gln Ile Asp Pro Thr Pro Val 405 410 415

Ile Val Thr Ser Asn Thr Asn Met Cys Ala Val Ile Asp Gly Asn Ser
420 425 430

Thr Thr Phe Glu His Gln Gln Pro Leu Gln Asp Arg Met Phe Lys Phe 435 440 445

Glu Leu Thr Arg Arg Leu Glu His Asp Phe Gly Lys Val Thr Lys Gln 450 460

Glu Val Lys Glu Phe Phe Arg Trp Ala Gln Asp His Val Thr Glu Val 465 470 475 480

Ala His Glu Phe Tyr Val Arg Lys Gly Gly Ala Asn Lys Arg Pro Ala 485 490 495

Pro Asp Asp Ala Asp Lys Ser Glu Pro Lys Arg Ala Cys Pro Ser Val 500 505 510

Ala Asp Pro Ser Thr Ser Asp Ala Glu Gly Ala Pro Val Asp Phe Ala 515 520 525

Asp Arg Tyr Gln Asn Lys Cys Ser Arg His Ala Gly Met Leu Gln Met 530 540

Leu Phe Pro Cys Lys Thr Cys Glu Arg Met Asn Gln Asn Phe Asn Ile 545 550 555 560

Cys Phe Thr His Gly Thr Arg Asp Cys Ser Glu Cys Phe Pro Gly Val 565 570 575

Ser Glu Ser Gln Pro Val Val Arg Lys Arg Thr Tyr Arg Lys Leu Cys 580 585 590

Ala Ile His His Leu Leu Gly Arg Ala Pro Glu Ile Ala Cys Ser Ala 595 600 605

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145-		•			150					155					160	
			-	ctg Leu 165	_	-									•	528
				aac Asn				-								576
				agc Ser			-									624
				gcg Ala		_							-	-		672
				Gly ggg												720
_				gag Glu 245	-	-	-									768
				tcc Ser	_		_	-		_	-		_		_	816
			-	acc Thr			_	Pro			_	-			_	864
_			-	att Ile			Asn	-			_		_		-	912
			-	cct Pro	-		-			_					-	960
_				ggg Gly 325						-						1008
	-		_	acc Thr					-		-					1056

\_ 340 345 350

														aat		1104
Phe	Tyr	_	Cys	Val	Asn	Trp		Asn	Glu	Asn	Phe		Phe	Asn	Asp	
		355				. •	360					365			•	
tgc	qtc	gac	aag	atg	qtq	atc	tgg	tgg	gag	gag	ggc	aag	atg	acg	gcc	1152
														Thr		
	370					375					380					
220	atc	ata	aaa	tcc	acc	220	acc	att	CT C	aac	aac	age	aan	gtg	cac	1200
-	_				-	_	_							Val		2200
385					390	-,-				395	,		-,-		400	
-	-		_	-	-	-		-	_		-			ccc		1248
Val	Asp	Gln	Lys		Lys	Ser	Ser	Ala		Ile	Asp	Pro	Thr	Pro	Val	
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atc	gtc	acc	tcc	aac	acc	aac	atg	tgc	gcc	gtg	att	gac	ggg	aac	agc	1296
Ile	Val	Thr	Ser	Asn	Thr	Asn	Met	Cys	Ala	Val	Ile	Asp	Gly	Asn	Ser	
			420					425					430			
					•											
					-	-	_	-	-	_		_		aaa		1344
Thr	Thr	435	GIU	HIS	GIN	GIN	440	Leu	GIN	Asp	Arg	445	Pne	Lys	Pne	
		433		•			440					113				
gaa	ctc	acc	cgc	cgt	ctg	gag	cat	gac	ttt	ggc	aag	gtg	aca	aag	cag	1392
Glu	Leu	Thr	Arg	Arg	Leu	Glu	His	Asp	Phe	Gly	Lys	Val	Thr	Lys	Gln	
	450					455					460					
																1440
•	_					-			_	-				gag	-	1440
465	val	гуѕ	GIU	Pne	470	Arg	irp	Ala	GIN	475	HIS	Val	1111	Glu	480	
403					470					475					400	
gcg	cat	gag	ttc	tac	gtc	aga	aag	ggt	gga	gcc	aac	aaa	aga	ccc	gcc	1488
Ala	His	Glu	Phe	Tyr	Val	Arg	Lys	Gly	Gly	Ala	Asn	Lys	Arg	Pro	Ala	
				485				,	490			•		495		
														<b>t</b> = -	~+ c	1536
	_					-								tca Ser		1336
PLO	Asp	Asp	500	Ash	гЛэ	261	GIU	505	Буз	Arg	AIa	Cys	510	Ser	Vai	
			500					505					010			
gcg	gat	cca	tcg	acg	tca	gac	gcg	gaa	gga	gct	ccg	gtg	gac	ttt	gcc	1584
Ala	Asp	Pro	Ser	Thr	Ser	Asp	Ala	Glu	Gly	Ala	Pro	Val	Asp	Phe	Ala	
		515					520					525		•		
						_		<b>.</b> .	_					<u> </u>		1622
-														gga Gl v		1632
wab	wrd	TAL	GTÀ	cys	wr.a	TTD	Leu	Ser	Set	AL 9	ne u	VIG	y	Gly	9411	

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Glu His Leu Pro Gly Ile Ser Asp Ser Phe Val Ser Trp Val Ala Glu 20 25 30

Lys Glu Trp Glu Leu Pro Pro Asp Ser Asp Met Asp Leu Asn Leu Ile 35 40 45

Glu Gln Ala Pro Leu Thr Val Ala Glu Lys Leu Gln Arg Asp Phe Leu
50 55 60

Val Gln Trp Arg Arg Val Ser Lys Ala Pro Glu Ala Leu Phe Phe Val 65 70 75 80

Gln Phe Glu Lys Gly Glu Ser Tyr Phe His Leu His Ile Leu Val Glu 85 90 95

Thr Thr Gly Val Lys Ser Met Val Leu Gly Arg Phe Leu Ser Gln Ile 100 105 110

Arg Asp Lys Leu Val Gln Thr Ile Tyr Arg Gly Ile Glu Pro Thr Leu 115 120 125

Pro Asn Trp Phe Ala Val Thr Lys Thr Arg Asn Gly Ala Gly Gly 130 135 140

Asn Lys Val Val Asp Glu Cys Tyr Ile Pro Asn Tyr Leu Leu Pro Lys 145 150 155 160

Thr Gln Pro Glu Leu Gln Trp Ala Trp Thr Asn Met Glu Glu Tyr Ile 165 170 175

Ser Ala Cys Leu Asn Leu Ala Glu Arg Lys Arg Leu Val Ala Gln His 180 185 190

PCT/US99/25694 WO 00/28061

Lew Thr His Val Ser Gln Thr Gln Glu Gln Asn Lys Glu Asn Leu Asn 195 200 Pro Asn Ser Asp Ala Pro Val Ile Arg Ser Lys Thr Ser Ala Arg Tyr 215 220 Met Glu Leu Val Gly Trp Leu Val Asp Arg Gly Ile Thr Ser Glu Lys 235 230 Gln Trp Ile Gln Glu Asp Gln Ala Ser Tyr Ile Ser Phe Asn Ala Ala 245 250 Ser Asn Ser Arg Ser Gln Ile Lys Ala Ala Leu Asp Asn Ala Gly Lys 260 265 Ile Met Ala Leu Thr Lys Ser Ala Pro Asp Tyr Leu Val Gly Pro Ala 28Û Pro Pro Ala Asp Ile Lys Thr Asn Arg Ile Tyr Arg Ile Leu Glu Leu . 295 Asn Gly Tyr Glu Pro Ala Tyr Ala Gly Ser Val Phe Leu Gly Trp Ala 310 315 . Gln Lys Arg Phe Gly Lys Arg Asn Thr Ile Trp Leu Phe Gly Pro Ala 330 325 Thr Thr Gly Lys Thr Asn Ile Ala Glu Ala Ile Ala His Ala Val Pro 340 345 350 Phe Tyr Gly Cys Val Asn Trp Thr Asn Glu Asn Phe Pro Phe Asn Asp 355 360 Cys Val Asp Lys Met Val Ile Trp Trp Glu Glu Gly Lys Met Thr Ala 370 375 380 Lys Val Val Glu Ser Ala Lys Ala Ile Leu Gly Gly Ser Lys Val Arg 395 400 385 390 Val Asp Gln Lys Cys Lys Ser Ser Ala Gln Ile Asp Pro Thr Pro Val 405 410 415 Ile Val Thr Ser Asn Thr Asn Met Cys Ala Val Ile Asp Gly Asn Ser 425 430 420

435

Thr Thr Phe Glu His Gln Gln Pro Leu Gln Asp Arg Met Phe Lys Phe 440

445

Glu Leu Thr Arg Arg Leu Glu His Asp Phe Gly Lys Val Thr Lys Gln 460 455 450 Glu Val Lys Glu Phe Phe Arg Trp Ala Gln Asp His Val Thr Glu Val 475 470 465 Ala His Glu Phe Tyr Val Arg Lys Gly Gly Ala Asn Lys Arg Pro Ala 490 485 Pro Asp Asp Ala Asp Lys Ser Glu Pro Lys Arg Ala Cys Pro Ser Val 510 505 500 Ala Asp Pro Ser Thr Ser Asp Ala Glu Gly Ala Pro Val Asp Phe Ala 520 515 Asp Arg Tyr Gly Cys Arg Trp Leu Ser Ser Arg Leu Ala Arg Gly Gln 540 535 Pro Leu 545 <210> 8 <211> 1200 <212> DNA <213> AAV-1 <220> <221> CDS <222> (1)..(1197) <400> 8 atg gag ctg gtc ggg tgg ctg gtg gac cgg ggc atc acc tcc gag aag 48 Met Glu Leu Val Gly Trp Leu Val Asp Arg Gly Ile Thr Ser Glu Lys 10 5 cag tgg atc cag gag gac cag gcc tcg tac atc tcc ttc aac gcc gct Gln Trp Ile Gln Glu Asp Gln Ala Ser Tyr Ile Ser Phe Asn Ala Ala 25 20 tec aac teg egg tee cag ate aag gee get etg gae aat gee gge aag Ser Asn Ser Arg Ser Gln Ile Lys Ala Ala Leu Asp Asn Ala Gly Lys 40 35 atc atg gcg ctg acc aaa tcc gcg ccc gac tac ctg gta ggc ccc gct 192 Ile Met Ala Leu Thr Lys Ser Ala Pro Asp Tyr Leu Val Gly Pro Ala 60 • 55 50

_										gag Glu	240
							-			tgg Trp 95	288
										ccg Pro	336
										gtg Val	384
			-	-						aat Asn	432
										acg Thr	480
										gtg Val 175	528
	-									ccc Pro	576
	-									aac Asn	624
		Phe								aaa Lys	672
-	Leu		_					Lys		aag Lys	720
_	-									gag Glu 255	768

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gcg	cat	gag	ttc	tac	gtc	aga	aag	ggt	gga	gcc	aac	aaa	aga	ccc	gcc	816
Ala	His	Glu	Phe	Tyr	Val	Arg	Lys	Gly	Gly	Ala	Asn	Lys	Arg	Pro	Ala	
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										_:_						0.64
						agc			_		_	-			•	864
Pro	Asp	Asp	Ala	Asp	Lys	Ser	Glu	Pro	Lys	Arg	Ala	Cys	Pro	Ser	Val	
		275					280					285				
aca	gat	сса	tca	acα	tca	gac	aca	gaa	gga	act	cca	ata	σac	ttt	acc	912
				_		Asp	-	_		-			-		_	
714	290	110	501	****	Jer	-	AIG	GIU	Gry	AIU		Val	Asp	FILE	AIG	
	290					295					300					
gac	agg	tac	caa	aac	aaa	tgt	tct	cgt	cac	gcg	ggc	atg	ctt	cag	atg	960
Asp	Arg	Tyr	Gln	Asn	Lys	Cys	Ser	Arg	His	Ala	Gly	Met	Leu	Gln	Met	
305					310					315					320	
ata			+~~	225	202	* ~ ~	~~~				•••					1008
-			_	-		tgc		-	-		-					1008
Leu	Phe	Pro	Cys		Thr	Cys	Glu	Arg		Asn	GIn	Asn	Phe		Ile	
				325					330					335		
tgc	ttc	acg	cac	ggg	acg	aga	gac	tgt	tca	gag	tgc	ttc	ccc	ggc	gtg	1056
_		_		• • •	_	Arg	-	-			-					
			340	,				345			~,~		350	<b>-</b> -,		
			340					343					330			
tca	gaa	tct	caa	ccg	gtc	gtc	aga	aag	agg	acg	tat	cgg	aaa	ctc	tgt	1104
Ser	Glu	Ser	Gln	Pro	Val	Val	Arg	Lys	Arg	Thr	Tyr	Arg	Lys	Leu	Cys	
		355					360					365				
acc	att	cat	cat	cta	cta	ggg	caa	act	ccc	gag	att	act	tac	tica	acc	1152
																1102
ALA		HIS	HIS	Leu	Leu	Gly	Arg	Ата	PIO	GIU		АТА	Cys	ser	Ala	
	370					375					380					
tgc	gat	ctg	gtc	aac	gtg	gac	ctg	gat	gac	tgt	gtt	tct	gag	caa	taa	1200
Cvs	Asp	Leu	٧a١	Asn	Val	Asp	Leu	Asp	Asp	Cvs	Val	Ser	Glu	Glp		
385				1	390			₽		395	- 48 46					
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<211> 399

<212> PRT

<213> AAV-1

<400> 9

Met Glu Leu Val Gly Trp Leu Val Asp Arg Gly Ile Thr Ser Glu Lys

1 5 10 15

Gln Trp Ile Gln Glu Asp Gln Ala Ser Tyr Ile Ser Phe Asn Ala Ala

\_ 20 25 30

Ser Asn Ser Arg Ser Gln Ile Lys Ala Ala Leu Asp Asn Ala Gly Lys 35 40 45

Ile Met Ala Leu Thr Lys Ser Ala Pro Asp Tyr Leu Val Gly Pro Ala 50 55 60

Pro Pro Ala Asp Ile Lys Thr Asn Arg Ile Tyr Arg Ile Leu Glu Leu 65 70 75 80

Asn Gly Tyr Glu Pro Ala Tyr Ala Gly Ser Val Phe Leu Gly Trp Ala 85 90 95

Gln Lys Arg Phe Gly Lys Arg Asn Thr Ile Trp Leu Phe Gly Pro Ala 100 105 110

Thr Thr Gly Lys Thr Asn Ile Ala Glu Ala Ile Ala His Ala Val Pro 115 120 125

Phe Tyr Gly Cys Val Asn Trp Thr Asn Glu Asn Phe Pro Phe Asn Asp 130 135 140

Cys Val Asp Lys Met Val Ile Trp Trp Glu Glu Gly Lys Met Thr Ala 145 150 155 160

Lys Val Val Glu Ser Ala Lys Ala Ile Leu Gly Gly Ser Lys Val Arg 165 170 175

Val Asp Gln Lys Cys Lys Ser Ser Ala Gln Ile Asp Pro Thr Pro Val 180 185 190

Ile Val Thr Ser Asn Thr Asn Met Cys Ala Val Ile Asp Gly Asn Ser 195 200 205

Thr Thr Phe Glu His Gln Gln Pro Leu Gln Asp Arg Met Phe Lys Phe 210 215 220

Glu Leu Thr Arg Arg Leu Glu His Asp Phe Gly Lys Val Thr Lys Gln 225 230 235 240

Glu Val Lys Glu Phe Phe Arg Trp Ala Gln Asp His Val Thr Glu Val 245 250 255

Ala His Glu Phe Tyr Val Arg Lys Gly Gly Ala Asn Lys Arg Pro Ala 260 265 270

Pro Asp Asp Ala Asp Lys Ser Glu Pro Lys Arg Ala Cys Pro Ser Val

275.

280

285

Ala Asp Pro Ser Thr Ser Asp Ala Glu Gly Ala Pro Val Asp Phe Ala 290 295 300 .

Asp Arg Tyr Gln Asn Lys Cys Ser Arg His Ala Gly Met Leu Gln Met 305 310 315 320

Leu Phe Pro Cys Lys Thr Cys Glu Arg Met Asn Gln Asn Phe Asn Ile 325 330 335

Cys Phe Thr His Gly Thr Arg Asp Cys Ser Glu Cys Phe Pro Gly Val 340 345 350

Ser Glu Ser Gln Pro Val Val Arg Lys Arg Thr Tyr Arg Lys Leu Cys 355 360 365

Ala Ile His His Leu Leu Gly Arg Ala Pro Glu Ile Ala Cys Ser Ala 370 375 380

Cys Asp Leu Val Asn Val Asp Leu Asp Asp Cys Val Ser Glu Gln 385 390 395

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<211> 969

<212> DNA

<213> AAV-1

<220>

<221> CDS

<222> (1)..(966)

<220>

<221> misc\_feature

<222> (943)..(944)

<223> minor splice site

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Met Glu Leu Val Gly Trp Leu Val Asp Arg Gly Ile Thr Ser Glu Lys
1 5 10 15

cag tgg atc cag gag gac cag gcc tcg tac atc tcc ttc aac gcc gct 96 Gln Trp Ile Gln Glu Asp Gln Ala Ser Tyr Ile Ser Phe Asn Ala Ala 20 25 30

tcc aac tcg cgg tcc cag atc aag gcc gct ctg gac aat gcc ggc aag 144

Ser Asn Ser Arg Ser Gln Ile Lys Ala Ala Leu Asp Asn Ala Gly Lys 35 40 atc atg gcg ctg acc aaa tcc gcg ccc gac tac ctg gta ggc ccc gct Ile Met Ala Leu Thr Lys Ser Ala Pro Asp Tyr Leu Val Gly Pro Ala 50 60 · 55 ccg ccc gcg gac att aaa acc aac cgc atc tac cgc atc ctg gag ctg Pro Pro Ala Asp Ile Lys Thr Asn Arg Ile Tyr Arg Ile Leu Glu Leu 65 aac ggc tac gaa cct gcc tac gcc tgc tcc gtc ttt ctc ggc tgg gcc 288 Asn Gly Tyr Glu Pro Ala Tyr Ala Gly Ser Val Phe Leu Gly Trp Ala 85 90 cag aaa agg ttc ggg aag cgc aac acc atc tgg ctg ttt ggg ccg gcc 336 Gln Lys Arg Phe Gly Lys Arg Asn Thr Ile Trp Leu Phe Gly Pro Ala 100 105 acc acg ggc aag acc aac atc gcg gaa gcc atc gcc cac gcc gtg ccc 384 Thr Thr Gly Lys Thr Asn Ile Ala Glu Ala Ile Ala His Ala Val Pro 120 125 115 ttc tac ggc tgc gtc aac tgg acc aat gag aac ttt ccc ttc aat gat 432 Phe Tyr Gly Cys Val Asn Trp Thr Asn Glu Asn Phe Pro Phe Asn Asp 130 135 tgc gtc gac aag atg gtg atc tgg tgg gag gag ggc aag atg acg gcc Cys Val Asp Lys Met Val Ile Trp Trp Glu Glu Gly Lys Met Thr Ala 150 155 160 145 aag gtc gtg gag tcc gcc aag gcc att ctc ggc ggc agc aag gtg cgc Lys Val Val Glu Ser Ala Lys Ala Ile Leu Gly Gly Ser Lys Val Arg 165 170 gtg gac caa aag tgc aag tcg tcc gcc cag atc gac ccc acc ccc gtg Val Asp Gln Lys Cys Lys Ser Ser Ala Gln Ile Asp Pro Thr Pro Val 180 185 atc gtc acc tcc aac acc aac atg tgc gcc gtg att gac ggg aac agc Ile Val Thr Ser Asn Thr Asn Met Cys Ala Val Ile Asp Gly Asn Ser 205 195 200 acc acc ttc gag cac cag cag ccg ttg cag gac cgg atg ttc aaa ttt 672 Thr Thr Phe Glu His Gln Gln Pro Leu Gln Asp Arg Met Phe Lys Phe 215 220 qaa ctc acc cqc cqt ctq qaq cat qac ttt qgc aag qtg aca aag cag

Glu 225	Leu	Thr	Arg	Arg	Leu 230	Glu	His	Asp	Phe	Gly 235	Lys	Val	Thr	Lys	Gln 240	
_	gtc Val		-			_										768
	cat His				-	-	_		_				-		-	816
	gat Asp	_		_		_			_		-	_			-	864
	gat Asp 290		-	_		-		_		_	_		-		-	912
-	agg Arg			_	_					_	_	-	_			960
	ctc Leu	tga								•	•					969
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	0> 1: Glu		Val	G1 y 5	Trp	Leu	Val	Asp	Arg 10	Gly	Ile	Thr	Ser	Glu 15	Lys	
Gln	Trp	Ile	Gln 20	Glu	Asp	Gln	Ala	Ser 25	туr	Ile	Ser	Phe	Asn 30	Ala	Ala	
Ser	Asn	Ser 35	Arg	Ser	Gln	Ile	Lys 40	Ala	Ala	Leu	Asp	Asn 45	Ala	Gly	Lys	
Ile								_		_				-		
	Met 50	Ala	Leu	Thr	Lys	Ser 55	Ala	Pro	Asp	Tyr	Leu 60	Val	GIÀ	Pro	Ala	

Asa Gly Tyr Glu Pro Ala Tyr Ala Gly Ser Val Phe Leu Gly Trp Ala Gln Lys Arg Phe Gly Lys Arg Asn Thr Ile Trp Leu Phe Gly Pro Ala Thr Thr Gly Lys Thr Asn Ile Ala Glu Ala Ile Ala His Ala Val Pro Phe Tyr Gly Cys Val Asn Trp Thr Asn Glu Asn Phe Pro Phe Asn Asp Cys Val Asp Lys Met Val Ile Trp Trp Glu Glu Gly Lys Met Thr Ala Lys Val Val Glu Ser Ala Lys Ala Ile Leu Gly Gly Ser Lys Val Arg Val Asp Gln Lys Cys Lys Ser Ser Ala Gln Ile Asp Pro Thr Pro Val Ile Val Thr Ser Asn Thr Asn Met Cys Ala Val Ile Asp Gly Asn Ser Thr Thr Phe Glu His Gln Gln Pro Leu Gln Asp Arg Met Phe Lys Phe Glu Leu Thr Arg Arg Leu Glu His Asp Phe Gly Lys Val Thr Lys Gln Glu Val Lys Glu Phe Phe Arg Trp Ala Gln Asp His Val Thr Glu Val Ala His Glu Phe Tyr Val Arg Lys Gly Gly Ala Asn Lys Arg Pro Ala Pro Asp Asp Ala Asp Lys Ser Glu Pro Lys Arg Ala Cys Pro Ser Val Ala Asp Pro Ser Thr Ser Asp Ala Glu Gly Ala Pro Val Asp Phe Ala Asp Arg Tyr Gly Cys Arg Trp Leu Ser Ser Arg Leu Ala Arg Gly Gln

Pro Leu

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Pro- 145	Val	Glu	Gln	Ser	Pro 150	Gln	Glu	Pro	Asp	Ser 155	Ser	Ser	Gly	Ile	Gly 160	
-			_	cag Gln 165		-				Leu						528
	-		-	tca Ser	-		-						-			576
-			-	gct Ala						_	-					624
-		-	_	gac Asp												672
				cat His												720
		-		cgc Arg 245			-									768
	_			tcc Ser	-	-										816
				agc Ser												864
				tca Ser												912
				ccc Pro												960
-	_		-	acg Thr 325												1008
ctt	acc	agc	acg	gtt	caa	gtc	ttc	tcg	gac	tcg	gag	tac	cag	ctt	ccg	1056

Leu_	Thr	Ser	Thr 340	Val	Gln	Val	Phe	Ser 345	Asp	Ser	Glu	Туr	Gln 350	Leu	Pro	
			ggc Gly													1104
-			atg Met													1152
-		-	gtg Val		-											1200
	_	_	ctg Leu	-	_											1248
	-		cct Pro 420			_	-									1296
			aat Asn													1344
			cag Gln													1392
_	Gly		cca Pro													1440
			tat Tyr												Asn	1488
		-	aat Asn 500											Leu		1536
	-	_	tcc Ser										Ser			1584
gac	gac	gaa	gac	aag	ttc	ttt	ccc	atg	agc	ggt	gtc	atg	att	ttt	gga	1632

Asp	Asp 530	Glu	Asp	Lys	Phe	Phe 535	Pro	Met	Ser	Gly	Val 540	Met	Ile	Phe	Gly	
	gag Glu															1680
	gac Asp	-		-			_									1728
	Gly ggg															1776
	gga Gly	-			-											1824
-	aga Arg 610															1872
	gat Asp															1920
-	aac Asn															1968
	cct Pro															2016
	tac Tyr														cag Gln	2064
	gaa Glu 690	Asn										Tyr				2112
	gca Ala			_		_	-				_					2160
tat	act	gag	cct	cgc	ccc	att	ggc	acc	cgt	tac	ctt	acc	cgt	ccc	ctg	2208

Tyr Thr Glu Pro Arg Pro Ile Gly Thr Arg Tyr Leu Thr Arg Pro Leu
725 730 735

taa 2211

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<211> 736

<212> PRT

<213> AAV-1

<400> 13

Met Ala Ala Asp Gly Tyr Leu Pro Asp Trp Leu Glu Asp Asn Leu Ser

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Glu Gly Ile Arg Glu Trp Trp Asp Leu Lys Pro Gly Ala Pro Lys Pro 20 25 30

Lys Ala Asn Gln Gln Lys Gln Asp Asp Gly Arg Gly Leu Val Leu Pro 35 40 45

Gly Tyr Lys Tyr Leu Gly Pro Phe Asn Gly Leu Asp Lys Gly Glu Pro 50 55 60

Val Asn Ala Ala Asp Ala Ala Ala Leu Glu His Asp Lys Ala Tyr Asp 65 70 75 80

Gln Gln Leu Lys Ala Gly Asp Asn Pro Tyr Leu Arg Tyr Asn His Ala 85 90 95

Asp Ala Glu Phe Gln Glu Arg Leu Gln Glu Asp Thr Ser Phe Gly Gly 100 105 110

Asn Leu Gly Arg Ala Val Phe Gln Ala Lys Lys Arg Val Leu Glu Pro 115 120 125

Leu Gly Leu Val Glu Glu Gly Ala Lys Thr Ala Pro Gly Lys Lys Arg 130 135 140

Pro Val Glu Gln Ser Pro Gln Glu Pro Asp Ser Ser Ser Gly Ile Gly
145 150 155 160

Lys Thr Gly Gln Gln Pro Ala Lys Lys Arg Leu Asn Phe Gly Gln Thr
165 170 175

Gly Asp Ser Glu Ser Val Pro Asp Pro Gln Pro Leu Gly Glu Pro Pro 180 185 190

Ala Thr Pro Ala Ala Val Gly Pro Thr Thr Met Ala Ser Gly Gly Gly 195 200 205

- Ala Pro Met Ala Asp Asn Asn Glu Gly Ala Asp Gly Val Gly Asn 'Ala 210 215 220
- Ser Gly Asn Trp His Cys Asp Ser Thr Trp Leu Gly Asp Arg Val Ile 225 230 235 240
- Thr Thr Ser Thr Arg Thr Trp Ala Leu Pro Thr Tyr Asn Asn His Leu 245 250 255
- Tyr Lys Gln Ile Ser Ser Ala Ser Thr Gly Ala Ser Asn Asp Asn His 260 265 270
- Tyr Phe Gly Tyr Ser Thr Pro Trp Gly Tyr Phe Asp Phe Asn Arg Phe 275 280 285
- His Cys His Phe Ser Pro Arg Asp Trp Gln Arg Leu Ile Asn Asn Asn 290 295 300
- Trp Gly Phe Arg Pro Lys Arg Leu Asn Phe Lys Leu Phe Asn Ile Gln 305 310 315 320
- Val Lys Glu Val Thr Thr Asn Asp Gly Val Thr Thr Ile Ala Asn Asn 325 330 335
- Leu Thr Ser Thr Val Gln Val Phe Ser Asp Ser Glu Tyr Gln Leu Pro 340 345 350
- Tyr Val Leu Gly Ser Ala His Gln Gly Cys Leu Pro Pro Phe Pro Ala 355 360 365
- Asp Val Phe Met Ile Pro Gln Tyr Gly Tyr Leu Thr Leu Asn Asn Gly 370 375 380
- Ser Gln Ala Val Gly Arg Ser Ser Phe Tyr Cys Leu Glu Tyr Phe Pro 385 390 395 400
- Ser Gln Met Leu Arg Thr Gly Asn Asn Phe Thr Phe Ser Tyr Thr Phe 405 410 415
- Glu Glu Val Pro Phe His Ser Ser Tyr Ala His Ser Gln Ser Leu Asp 420 425 430
- Arg Leu Met Asn Pro Leu Ile Asp Gln Tyr Leu Tyr Tyr Leu Asn Arg
  435
  440
  445

Thr\_Gln Asn Gln Ser Gly Ser Ala Gln Asn Lys Asp Leu Leu Phe Ser Arg Gly Ser Pro Ala Gly Met Ser Val Gln Pro Lys Asn Trp Leu Pro Gly Pro Cys Tyr Arg Gln Gln Arg Val Ser Lys Thr Lys Thr Asp Asn Asn Asn Ser Asn Phe Thr Trp Thr Gly Ala Ser Lys Tyr Asn Leu Asn Gly Arg Glu Ser Ile Ile Asn Pro Gly Thr Ala Met Ala Ser His Lys Asp Asp Glu Asp Lys Phe Phe Pro Met Ser Gly Val Met Ile Phe Gly Lys Glu Ser Ala Gly Ala Ser Asn Thr Ala Leu Asp Asn Val Met Ile Thr Asp Glu Glu Glu Ile Lys Ala Thr Asn Pro Val Ala Thr Glu Arg Phe Gly Thr Val Ala Val Asn Phe Gln Ser Ser Ser Thr Asp Pro Ala Thr Gly Asp Val His Ala Met Gly Ala Leu Pro Gly Met Val Trp Gln Asp Arg Asp Val Tyr Leu Gln Gly Pro Ile Trp Ala Lys Ile Pro His €15 Thr Asp Gly His Phe His Pro Ser Pro Leu Met Gly Gly Phe Gly Leu Lys Asn Pro Pro Pro Gln Ile Leu Ile Lys Asn Thr Pro Val Pro Ala Asn Pro Pro Ala Glu Phe Ser Ala Thr Lys Phe Ala Ser Phe Ile Thr 

Gln Tyr Ser Thr Gly Gln Val Ser Val Glu Ile Glu Trp Glu Leu Gln 675 680 685

Lys Glu Asn Ser Lys Arg Trp Asn Pro Glu Val Gln Tyr Thr Ser Asn

Tyr Ala Lys Ser Ala Asn Val Asp Phe Thr Val Asp Asn Asn Gly Leu 705 710 715 720

Tyr Thr Glu Pro Arg Pro Ile Gly Thr Arg Tyr Leu Thr Arg Pro Leu
725 730 735

<210> 14

<211> 1800

<212> DNA

<213> AAV-1

<220>

<221> CDS

<222> (1)..(1797)

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Thr Ala Pro Gly Lys Lys Arg Pro Val Glu Gln Ser Pro Gln Glu Pro

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gac tcc tcc tcg ggc atc ggc aag aca ggc cag cag ccc gct aaa aag 96
Asp Ser Ser Ser Gly Ile Gly Lys Thr Gly Gln Gln Pro Ala Lys Lys
20 25 30

aga ctc aat ttt ggt cag act ggc gac tca gag tca gtc ccc gat cca 144 Arg Leu Asn Phe Gly Gln Thr Gly Asp Ser Glu Ser Val Pro Asp Pro 35 40 45

caa cct ctc gga gaa cct cca gca acc ccc gct gct gtg gga cct act 192
Gln Pro Leu Gly Glu Pro Pro Ala Thr Pro Ala Ala Val Gly Pro Thr
50 55 60

aca atg gct tca ggc ggt ggc gca cca atg gca gac aat aac gaa ggc 240
Thr Met Ala Ser Gly Gly Gly Ala Pro Met Ala Asp Asn Asn Glu Gly
65 70 75 80

gcc gac gga gtg ggt aat gcc tca gga aat tgg cat tgc gat tcc aca 288
Ala Asp Gly Val Gly Asn Ala Ser Gly Asn Trp His Cys Asp Ser Thr
85 90 95

tgg ctg ggc gac aga gtc atc acc acc agc acc cgc acc tgg gcc ttg 336
Trp Leu Gly Asp Arg Val Ile Thr Thr Ser Thr Arg Thr Trp Ala Leu
100 105 110

ccc acc tac aat aac cac ctc tac aag caa atc tcc agt gct tca acg

Pro Thr Tyr Asn Asn His Leu Tyr Lys Gln Ile Ser Ser Ala Ser Thr

115

120

125

	-				aac Asn											432
		-			aga Arg 150											480
_					aac Asn											528
					atc Ile											576
-				_	aat Asn											624
-	_			_	ctt Leu											672
_					ccg Pro 230											720
	_	_			aat Asn	-				Val						768
Tyr	Cys	Leu	Glu 260	Туr	ttc Phe	Pro	Ser	Gln 265	Met	Leu	Arg	Thr	Gly 270	Asn	Asn	816
Phe	Thr	Phe 275	Ser	Tyr		Phe	Glu 280	Glu	Val	Pro	Phe	His 285	Ser	Ser	Tyr	864
Ala	His 290	Ser	Gln	Ser	Leu	Asp 295	Arg	Leu	Met	Asn	9ro 300	Leu	Ile	Asp	caa Gln	912
	Leu			_							Ser				caa Gln 320	960

				ctg Leu 325										Val	1008
_				tgg Trp											1056
				aca Thr	-										1104
_				aac Asn											1152
	-			tca Ser											1200
		-	-	att Ile 405	Phe		•								1248
-	-	_		gtc Val	-										1296
			_	acc Thr											1344
-	_	Ser		gac Asp		 Thr		-	-		Ala	_		_	1392
	Pro		_	gtg Val						Tyr					1440
				att Ile 485					His						1488
	-			ttt Phe									Leu		1536

1584 aaa aac acg cct gtt cct gcg aat cct ccg gcg gag ttt tca gct aca Lys Asn Thr Pro Val Pro Ala Asn Pro Pro Ala Glu Phe Ser Ala Thr 515 525 520 aag ttt gct tca ttc atc acc caa tac tcc aca gga caa gtg agt gtg 1632 Lys Phe Ala Ser Phe Ile Thr Gln Tyr Ser Thr Gly Gln Val Ser Val 540 530 535 1680 gaa att gaa tgg gag ctg cag aaa gaa aac agc aag cgc tgg aat ccc Glu Ile Glu Trp Glu Leu Gln Lys Glu Asn Ser Lys Arg Trp Asn Pro 545 550 555 gaa gtg cag tac aca tcc aat tat gca aaa tct gcc aac gtt gat ttt 1728 Glu Val Gln Tyr Thr Ser Asn Tyr Ala Lys Ser Ala Asn Val Asp Phe 570 575 565 act gtg gac aac aat gga ctt tat act gag cct cgc ccc att ggc acc 1776 Thr Val Asp Asn Asn Gly Leu Tyr Thr Glu Pro Arg Pro Ile Gly Thr 580 585 1800 cgt tac ctt acc cgt ccc ctg taa Arg Tyr Leu Thr Arg Pro Leu 595 <210> 15 <211> 599 <212> PRT <213> AAV-1 <400> 15 Thr Ala Pro Gly Lys Lys Arg Pro Val Glu Gln Ser Pro Gln Glu Pro 10 1 Asp Ser Ser Ser Gly Ile Gly Lys Thr Gly Gln Gln Pro Ala Lys Lys 25 20 Arg Leu Asn Phe Gly Gln Thr Gly Asp Ser Glu Ser Val Pro Asp Pro 35 40 Gln Pro Leu Gly Glu Pro Pro Ala Thr Pro Ala Ala Val Gly Pro Thr 55 Thr Met Ala Ser Gly Gly Gly Ala Pro Met Ala Asp Asn Asn Glu Gly 70 75

Ala Asp Gly Val Gly Asn Ala Ser Gly Asn Trp His Cys Asp Ser Thr

_															
Trp	Leu	Gly	Asp 100	Arg	Val	Ile	Thr	Thr 105	Ser	Thr	Arg	Thr	Trp 110	Ala	Leu
Pro	Thr	туr 115	Asn	Äsn	His	Leu	Tyr 120	Lys	Gln	Ile	Ser	Ser 125	Ala	Ser	Thr
Gly	Ala 130	Ser	Asn	Asp	Asn	His 135	Tyr	Phe	Gly	Tyr	Ser 140	Thr	Pro	Trp	Gly
туг 145	Phe	Asp	Phe	Asn	Arg 150	Phe	His	Cys	His	Phe 155	Ser	Pro	Arg	Asp	Trp 160
Gln	Arg	Leu	Ile	Asn 165	Asn	Asn	Trp	Gly	Phe 170	Arg	Pro	Lys	Arg	Leu 175	Asn
Phe	Lys	Leu	Phe 180	Asn	Ilė	Gln	Val	Lys 185	Glu	Val	Thr	Thr	Asn 190	Asp	Gly
Val	Thr	Thr 195	Ile	Ala	Asn	Asn	Leu 200	Thr	Ser	Thr	Val	Gln 205	Val	Phe	Ser
Asp	Ser 210	Glu	Tyr	Gln	Leu	Pro 215	Tyr	Val	Leu	Gly	Ser 220	Ala	His	Gln	Ğlу
Cys 225	Leu	Pro	Pro	Phe	Pro 230	Ala	Asp	Val	Phe	Met 235	Ile	Pro	Gln	Tyr	Gly 240
Туr	Leu	Thr	Leu	Asn 245	Asn	Gly	Ser	Gln	Ala 250	Val	Gly	Arg	Ser	Ser 255	Phe
Туr	Cys	Leu	Glu 260	-	Phe	Pro	Ser	Gln 265	Met	Leu	Arg	Thr	Gly 270	Asn	Asn
Phe	Thr	Phe 275		Tyr	Thr	. Phe		Glu		Pro	Phe	200	Ser	Ser	Tyr
Ala	His 290		Gln	Ser	Leu	Asp 295		Leu	Met	Asn	Pro 300	Leu	Ile	Asp	Gln
Туг 305		Tyr	Tyr	Leu	Asn 310	Arg	Thr	Gln	Asn	Gln 315		Gly	Ser	Ala	Gln 320
Asn	Lys	Asp	Leu	Leu 325	Phe	Ser	Arg	Gly	ser 330		Ala	Gly	Met	Ser 335	
Gln	Pro	Lys	Asn	Trp	Leu	Pro	Gly	Pro	Cys	Tyr	Arg	Gln	Gln	Arg	Val

Ser Lys Thr Lys Thr Asp Asn Asn Ser Asn Phe Thr Trp Thr Gly Ala Ser Lys Tyr Asn Leu Asn Gly Arg Glu Ser Ile Ile Asn Pro Gly Thr Ala Met Ala Ser His Lys Asp Asp Glu Asp Lys Phe Phe Pro Met Ser Gly Val Met Ile Phe Gly Lys Glu Ser Ala Gly Ala Ser Asn Thr 410 . Ala Leu Asp Asn Val Met Ile Thr Asp Glu Glu Glu Ile Lys Ala Thr Asn Pro Val Ala Thr Glu Arg Phe Gly Thr Val Ala Val Asn Phe Gln Ser Ser Ser Thr Asp Pro Ala Thr Gly Asp Val His Ala Met Gly Ala Leu Pro Gly Met Val Trp Gln Asp Arg Asp Val Tyr Leu Gln Gly Pro Ile Trp Ala Lys Ile Pro His Thr Asp Gly His Phe His Pro Ser Pro Leu Met Gly Gly Phe Gly Leu Lys Asn Pro Pro Pro Gln Ile Leu Ile Lys Asn Thr Pro Val Pro Ala Asn Pro Pro Ala Glu Phe Ser Ala Thr Lys Phe Ala Ser Phe Ile Thr Gln Tyr Ser Thr Gly Gln Val Ser Val 5.30 Glu Ile Glu Trp Glu Leu Gln Lys Glu Asn Ser Lys Arg Trp Asn Pro Glu Val Gln Tyr Thr Ser Asn Tyr Ala Lys Ser Ala Asn Val Asp Phe 

Arg Tyr Leu Thr Arg Pro Leu

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Phe Asp Phe Asn Arg Phe His Cys His Phe Ser Pro Arg Asp Trp Gln 85 90 95

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Lys Leu Phe Asn Ile Gln Val Lys Glu Val Thr Thr Asn Asp Gly Val 115 120 125

Thr Thr Ile Ala Asn Asn Leu Thr Ser Thr Val Gln Val Phe Ser Asp 130 135 140

Ser Glu Tyr Gln Leu Pro Tyr Val Leu Gly Ser Ala His Gln Gly Cys 145 150 155 160

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16

Int. Jonal Application No PCT/US 99/25694

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CLASSIF PC 7	-C12N15/86 C12N15/35 C12N5/	/10 A61K48/00				
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- DOCUM	ENTS CONSIDERED TO BE RELEVANT					
Category *	Citation of document, with indication, where appropriate, of the	ne relevant passages	Relevant to claim No.			
1	RUTLEDGE E. A. ET AL.: "Infecand vectors derived from adeno virus (AAV) serotypes other th	-associated	1-23			
	2." JOURNAL OF VIROLOGY, vol. 72, no. 1, January 1998 ( pages 309-319, XP002137089 ISSN: 0022-538X cited in the application the whole document	1998-01),				
Y	WO 98 11244 A (SAFER BRIAN ;US (US); CHIORINI JOHN A (US); KC M) 19 March 1998 (1998-03-19) the whole document		1-23			
		-/				
X Fur	rther documents are listed in the continuation of box C.	Patent family members are listed	d In annèx.			
"A" docurr cone "E" earlier filing "L" docurr which citati "O" docurr other "P" docurr	nent defining the general state of the art which is not idered to be of particular relevance if document but published on or after the international judate nent which may throw doubts on priority claim(s) or the cited to establish the publication date of another ion or other special reason (as epecified) ment referring to an oral disclosure, use, exhibition or in means are the priority date of the international filing date but then the priority date claimed	or priority date and not in conflict with cited to understand the principle or the invention.  "X" document of particular relevance; the cannot be considered novel or cannot involve an inventive step when the decument of particular relevance; the cannot be considered to involve an indecument is combined with one or n ments, such combination being obvin the art.	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person aktiled			
	e actual completion of the international search	Date of mailing of the International s	earch report			
	8 May 2000	22/05/2000				
	d mailing address of the ISA	Authorized officer				

## · · · · INTERNATIONAL SELLACH REPORT

Int Honal Application No PCT/US 99/25694

		PCT/US 99/25694
C.(Continue Category *	ction) DOCUMENTS CONSIDERED TO BE RELEVANT  Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	XIAO W. ET AL.: "Gene therapy vectors based on adeno-associated virus type 1." JOURNAL OF VIROLOGY,	1–23
	vol. 73, no. 5, May 1999 (1999-05), pages 3994-4003, XP002137090 ISSN: 0022-538X the whole document	•
	·	
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## INTERNATIONAL SEARCH REPORT

...emational application No.

PCT/US 99/25694

Boxi	Observations where certain claims were found unsearchable (Continuation of Item 1 of Tirst sneet)
This Int	emational Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. X	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:  Remark: Although claims 18-20 and 22, as far as an in vivo application is concerned, are directed to a method of treatment of the human or animal body, the search has been carried out and based on the alleged effects of the compound/composition.  Claims Nos.:
£ [	because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II	Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)
This In	ternational Searching Authority found multiple inventions in this international application, as follows:
1.	As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2.	As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. [	No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Rema	The additional search fees were accompanied by the applicant's protest.  No protest accompanied the payment of additional search fees.

-INTERNATIONAL SEALCH REPORT

Information on patent family members

int Bonal Application No PCT/US 99/25694

Patent document cited in search report	Publication date	Patent family member(s)	Publication date		
WO 9811244 A	19-03-1998	AU 4645697 A EP 0932694 A	02-04-1998 04-08-1999		